

In-vitro* Antitumor Activity and Antifungal Activity of Pennogenin Steroidal Saponins from *paris Polyphylla* var. *yunnanensis

Liancai Zhu^{a*}, Jun Tan^{a, b}, Bochu Wang^{a*}, Luhan Guan^a, Yuping Liu^c and Chao Zheng^a

^aKey Laboratory of Biorheological Science and Technology (Chongqing University), Ministry of Education, Bioengineering College, Chongqing University, Chongqing 400044, China. ^bDepartment of Life Science & Chemistry, Chongqing Education College, Chongqing, 400067, China. ^cChemistry and Chemical Engineering College, Chongqing University, Chongqing, 400030, China.

Abstract

Paris polyphylla Smith var. *yunnanensis*, has been used in traditional Chinese medicine for its antibiotic and anti-inflammatory properties; in addition it has been used to cure liver cancer in particular. In this current study, β -ecdysterone (1) and three pennogenin steroidal saponins (2-4) were isolated from the EtOH extract of *Paris polyphylla* var. *yunnanensis*, and then tested for their antitumor and antifungal activities. Spectroscopic data was used to confirm their structures. Their antitumor properties were determined by using an MTT assay in addition to ethidium bromide and acridine orange staining techniques. Compounds 2, 3 and 4 exhibited significant anti-proliferation activities against HepG2 cells, with IC₅₀ values of 13.5 μ M, 9.7 μ M and 11.6 μ M respectively, obtained following 48 h treatment. Furthermore, we found these pennogenin steroidal saponins could induce HepG2 cells apoptosis at a concentration of 20 μ M after 48 h treatment. Compounds 2, 3 and 4 were confirmed to exhibit moderate antifungal activity. The minimum inhibitory concentration (MIC) of compounds 2, 3 and 4 against *saccharomyces cerevisiae hansen* were 2.5 mg.mL⁻¹, 0.6 mg.mL⁻¹ and 0.6 mg.mL⁻¹, respectively. The MIC of compounds 2, 3 and 4 against *Candida albicans* were 1.2 mg.mL⁻¹, 0.6 mg.mL⁻¹ and 1.2 mg.mL⁻¹, respectively. The analysis of the bioactivity-structure relationship shows that the sugar moiety plays a critical role in the activity of steroid moiety. Our results suggest that these three pennogenin steroidal saponins could be utilized to develop anticancer medicines.

Keywords: *Paris polyphylla* var. *yunnanensis*; Pennogenin steroidal saponins; Antitumor activity; Antifungal activity.

Introduction

Medical herbs have been used for a few thousands years and are considered to be one of the most promising sources of new medicines and leading compounds due to their

therapeutic effects demonstrated in clinical use. *Paris polyphylla* Smith var. *yunnanensis*, used as a Chinese traditional medicine, is widely distributed in China. It has been widely used due to its antibiotic and anti-inflammatory properties. In addition this herbal medicine has been used to treat liver cancer in particular (1). Certain steroidal saponins isolated from *paris polyphylla* have been reported to convey antitumor and

* Corresponding author:

E-mail: zhuliancai75@126.com

immuno-stimulating properties (2-4). This paper reports the isolation, structure determination and biological activities of β -ecdysterone and three pennogenin steroidal saponins (2-4). We found that compounds 2-4 exhibited significant anti-proliferation activities against HepG2 cells and induced apoptosis of HepG2 cells.

Experimental

Isolation

The dried aerial part of *Paris polyphylla* var. *yunnanensis*, purchased from the Western Medicine City of Chongqing, China, were twice extracted with 10 volumes of EtOH under reflux at 8°C for 60 min. The filtrate was combined and evaporated to achieve the ethanol extract. The ethanol extract was partitioned successively with chloroform, ethyl acetate and *n*-butanol.

The *n*-butanol soluble fraction that showed major inhibitory activity on HepG2 cells, was subjected to fractionation using silica gel (Qingdao Ocean Chemical Company, China) and column eluted with CHCl₃/MeOH (10 : 2-6 : 2 gradients) resulting in the formation of four fractions (F1-F4). F2 was then subjected to fractionation using silica gel column eluted with CHCl₃/MeOH (12 : 2-10 : 2 gradients) to afford Compound 1. F3 was subjected to fractionation using Source 15 RPC (GE Healthcare Life Sciences) column eluted with MeOH/H₂O (3 : 7-10 : 0 gradients) to afford compounds 2-4.

Data of isolated compounds

The spectra data of compounds 1-4 were measured: NMR on a 500 MHz Bruker DRX-500 instrument (Bruker, German) and MS on a VG Auto-spec3000 mass spectrometer (VG, England).

β -ecdysterone

White needle crystal; ESI-MS m/z : 479 [M - H]⁻; ¹H-NMR(500MHz, DMSO-d₆): δ_H 0.74 (3H, s, H-19), δ_H 0.82 (3H, s, H-18), δ_H 1.04 (3H, s, H-21), δ_H 1.06 (6H, s, H-26, H-27), δ_H 5.65 (1H, s, H-7), δ_H 3.13 (1H, d, J = 9.8Hz, H-22), δ_H 3.05 (1H, t, H-9), δ_H 4.39 (1H, m, H-2), δ_H 4.51 (1H, br s, H-3); ¹³C NMR (125 MHz, DMSO-d₆) Table 1.

pennogenin-3-O- α -L-rhamnopyranosyl (1→2)- β -D-glucopyranoside

Colorless needle crystal; ESI-MS m/z : 737 [M - H]⁻; ¹H-NMR(500MHz, DMSO-d₆): δ_H 0.68 (3H, d, J = 6.0 Hz, H-27), δ_H 0.98 (3H, s, H-18), δ_H 1.13 (3H, s, H-19), δ_H 1.23 (3H, d, J = 6.8 Hz, H-21), δ_H 1.86 (3H, d, J = 7.7 Hz, rha H-6'), δ_H 3.55 (2H, m, H-26), δ_H 3.82 (1H, m, H-3), δ_H 5.00 (1H, d, J = 4.5 Hz, glc H-1), δ_H 5.24 (1H, s, H-6), δ_H 6.37 (1H, s, rha H-1'); ¹³C NMR (125 MHz, DMSO-d₆) Table 1.

pennogenin-3-O- α -L-rhamnopyranosyl (1→4)- α -L-rhamnopyranosyl

(1→4)-[α -L-rhamnopyranosyl (1→2)]- β -D-glucopyranoside.

Colorless needle crystal; ESI-MS m/z : 1029 [M-H]⁻; ¹H-NMR(500MHz, DMSO-d₆): δ_H 0.69 (3H, d, J = 5.7 Hz, H-27), δ_H 0.94 (3H, s, H-18), δ_H 1.08 (3H, s, H-19), δ_H 1.22 (3H, d, J = 7.4 Hz, H-21), δ_H 1.58 (3H, d, J = 6.8 Hz, rha H-6'), δ_H 1.60 (3H, d, J = 6.9 Hz, rha H-6''), δ_H 1.86 (3H, d, J = 7.7 Hz, rha H-6'''), δ_H 3.60 (2H, m, H-26), δ_H 3.85 (1H, m, H-3), δ_H 4.98 (1H, d, J = 4.5 Hz, glc H-1), δ_H 5.24 (1H, s, H-6), δ_H 5.78 (1H, s, rha H-1'), δ_H 6.27 (1H, s, rha H-1''), δ_H 6.35 (1H, s, rha H-1'''); ¹³C NMR (125 MHz, DMSO-d₆) Table 1.

24- α -hydroxyl-pennogenin-3-O- α -L-rhamnopyranosyl (1→2)-[α -L-arabinofuranosyl (1→4)]- β -D-glucopyranoside.

Colorless needle crystal. ESI-MS m/z : 885 [M-H]⁻; ¹H-NMR(500MHz, DMSO-d₆): δ_H 0.70 (3H, d, J = 5.7 Hz, H-27), δ_H 0.94 (3H, s, H-18), δ_H 1.07 (3H, s, H-19), δ_H 1.22 (3H, d, J = 6.9 Hz, H-21), δ_H 1.58 (3H, d, J = 6.0 Hz, rha H-6'), δ_H 3.52 (2H, m, H-26), δ_H 3.83 (1H, m, H-3), δ_H 4.98 (1H, d, J = 4.5 Hz, glc H-1), δ_H 5.30 (1H, s, H-6), δ_H 5.82 (1H, s, ara H-1), δ_H 6.25 (1H, s, rha H-1); ¹³C NMR (125 MHz, DMSO-d₆) Table 1.

Assay for antitumor activity

Cell culture

HepG2 cells were cultured in RPMI 1640 medium (HyClone, USA) supplemented with 10% fetal bovine serum (HyClone, USA), 100 units/mL penicillin, 100 μ g/mL streptomycin, L-glutamine (0.03%, w/v) and sodium bicarbonate (2.2%, w/v). The cell cultures were

Table 1. The ^{13}C -NMR chemical shifts of compounds 1-4 (DMSO- d_6 , δ).

Aglycone moiety	Compound				Sugar moiety	Compound		
	1	2	3	4		2	3	4
1	36.7	37.4	37.7	36.9		glc	glc	glc
2	66.9	30.2	30.9	30.9	1	98.7	98.2	100.1
3	66.7	78.2	77.9	78.1	2	76.9	76.6	78.0
4	31.0	38.1	37.7	37.7	3	76.4	75.8	75.4
5	50.2	140.8	140.3	140.3	4	71.0	76.2	76.3
6	203.0	121.8	121.4	121.4	5	76.6	76.0	76.0
7	120.6	31.3	31.3	31.3	6	63.4	63.0	63.3
8	165.5	31.8	31.4	31.4		rha	rha	rha
9	33.3	50.0	49.6	49.6	1	100.5	107.8	100.4
10	37.8	36.8	36.9	36.4	2	72.3	72.5	72.5
11	20.4	20.5	20.1	20.1	3	72.8	70.5	71.3
12	31.7	31.7	31.6	31.4	4	72.6	81.3	71.9
13	50.2	44.1	43.7	43.7	5	68.4	68.0	68.0
14	83.1	52.4	52.0	52.0	6	18.2	17.2	18.2
15	30.0	32.0	31.6	31.6			rha	ara
16	20.4	88.8	88.4	88.4	1		100.5	101.1
17	48.8	89.4	88.9	89.0	2		72.0	82.5
18	17.3	17.5	17.2	17.2	3		72.4	78.3
19	21.1	19.5	19.1	19.0	4		75.8	86.9
20	75.9	44.8	44.4	44.4	5		68.5	61.3
21	20.2	9.8	9.4	9.4	6		17.8	
22	75.9	109.2	108.8	108.8			rha	
23	24.0	30.6	30.9	30.9	1		100.2	
24	41.5	29.2	28.1	63.0	2		71.9	
25	75.8	29.5	29.1	29.1	3		70.5	
26	26.2	66.3	65.9	65.9	4		72.1	
27	29.1	17.0	16.8	16.8	5		68.5	
					6		16.8	

kept in a humidified incubator containing 5% CO_2 set at 37°C. Subcultures were performed with 0.05% trypsin and 0.02% EDTA in phosphate-buffered saline solution (Gibco BRL Co., USA).

MTT assay

The MTT assay was performed according to the method set out by Mosmann(5). The HepG2 cells were plated into 96-well microtiter plates at a density of 1×10^4 cells/well. After 24 h, the culture medium was replaced with 200 μL RPMI 1640 medium supplemented with 10% fetal bovine serum containing varying concentrations (0, 5, 10, 20, 40 and 80 μM) of compound 2, 3 and

4. The cells were then subjected to incubation for 24 and 48 h. The final concentration of solvent was less than 0.1% in the cell culture medium. The culture solutions were removed and replaced with 90 μL of culture medium. Ten microliters of sterile filtered MTT (Sigma, USA) solution (5 mg/mL) suspended in PBS (pH = 7.4) was added to each well to achieve a final concentration of 0.5 mg MTT/mL. The cells were then incubated at 37°C for 4 h. After the medium and unreacted dye was removed, 200 μL of DMSO was added to each well. The absorbance at 490 nm of the dissolved solution was measured using a Bio-Rad 680 microplate reader (BIORAD, USA).

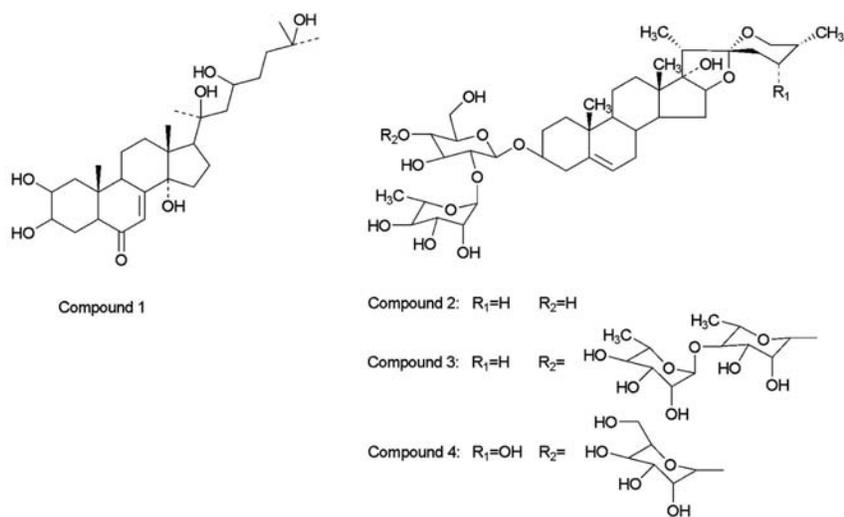


Figure 1. The structures of compound 1, 2, 3 and 4 from the EtOH extract of *paris polyphylla* var. *yunnanensis*.

The relative cell viability (%) of the control wells containing cell culture medium without the tested compound was calculated by dividing the absorbance of treated cells by that of the controls in each experiment. The IC_{50} was calculated as the tested compound concentration by means of SPSS statistical software, which inhibits the growth of 50% of cells in relation to non-treated control cells.

AO/EB staining assay

The cells were cultured on coverslips and kept in a 60 Petri dish for 24 h before treatment. Following treatment for 48 h with compounds 2, 3 and 4 at a concentration of 20 μ M, the cells without fixation were loaded with 100 μ L of freshly-prepared AO/EB (Sigma, USA) staining solution (100 μ g/mL). The cells were then immediately (less than 20 mins) observed under a fluorescence microscope (OLYMPUS, JAPAN).

Assay for antifungal activity

The antifungal activity against *saccharomyces cerevisiae hansen* and *candida albicans* of compounds 1-4 was evaluated by determining their minimum inhibitory concentrations (MIC), using broth microdilution techniques. The MIC values were determined in RPMI-

1640 (Hyclone, USA). Stock solutions of pure compounds were twofold diluted with RPMI-1640 from 20.0 to 0.6 mg/mL and aliquoted into test tubes. Each tube was then inoculated with 25 μ L of a standardized solution containing 10⁶ cfu/mL fungi suspended in sterile NaCl solution. After 24 h of incubation at 37°, the MICs were determined by using the optical density of the solutions. The lowest concentration of drug that inhibited all fungal growth was determined as being the MIC.

Results and Discussion

Isolation and identification

The EtOH extract of *paris polyphylla* var. *yunnanensis* was suspended in H₂O and fractionated using chloroform, ethyl acetate and *n*-butanol, successively. The *n*-butanol soluble fraction, which showed major inhibitory activity on HepG2 cells, was repeatedly subjected to silica gel column chromatography and Source 15 RPC column chromatography to afford compounds 1-4. The structures of compounds 1-4 were analyzed using ¹H, ¹³C NMR and MS. Finally, the structures were confirmed upon comparison with reference data. They were identified as β -ecdysterone(1) [6], pennogenin-3-O- α -L-rhamnopyranosyl (1 \rightarrow 2)- β -D-

Table 2. Antifungal activity of compounds 1, 2, 3 and 4.

Strains	MIC (mg.mL ⁻¹)			
	Compound 1	Compound 2	Compound 3	Compound 4
<i>Saccharomyces cerevisiae hansen</i>	20.0	2.5	0.6	0.6
<i>Candida albicans</i>	10.0	1.2	0.6	1.2

glucopyranoside(2) [7], pennogenin-3-O- α -L-rhamnopyranosyl (1 \rightarrow 4)- α -L-rhamnopyranosyl (1 \rightarrow 4)-[α -L-rhamnopyranosyl (1 \rightarrow 2)]- β -D-glucopyranoside(3) [7-8], and 24- α -hydroxyl-pennogenin-3-O- α -L-rhamnopyranosyl (1 \rightarrow 2)-[α -L-arabinofuranosyl (1 \rightarrow 4)]- β -D-glucopyranoside(4) [9], as shown in Figure 1. The last three are pennogenin steroidal saponins.

Antitumor activity

Two experiments were carried out to determine the antitumor activity of the identified compounds.

The potential anti-proliferative effect of these identified compounds on the viability of HepG2 cells was evaluated by means of the MTT assay. Compound 1 showed no significant effect on inhibiting the growth and proliferation of HepG2 cells (data not shown). Cell viability is expressed as the mean percentage (\pm SD) of viable cells in comparison with untreated cells (taken as 100% viable) for different concentrations of compounds 2, 3 and 4. These compounds significantly inhibited the growth and proliferation of HepG2 cells in a dose- and time-dependent manner (Figure 2). The cell viability was shown to be 12.32%, 6.48% and 9.6% following treatment with 20 μ M of compounds 2, 3 and 4 for 48 h, respectively. The estimated IC₅₀ (50% of growth is inhibited) value of compounds 2, 3 and 4 obtained for 48 h treatment were 13.5 μ M, 9.7 μ M and 11.6 μ M respectively, analyzed by SPSS software.

To further address the cell death pattern, HepG2 cells after treatment for 48 h with compounds 2, 3 and 4 at a concentration of 20 μ M were stained with the acridine orange and ethidium bromide (AO/EB), and immediately observed under a fluorescence microscope. AO/EB staining combines the differential uptake of fluorescent DNA binding dyes AO and EB and the morphologic aspect of chromatin condensation within the stained nucleus. This

allows viable, apoptotic and necrotic cells to be distinguished from each another. Viable cells possess a uniformly bright green nucleus. Early apoptotic cells show bright green areas of condensed or fragmented chromatin within the nucleus. Necrotic cells exhibit a uniformly bright orange nucleus. After staining with AO/EB, the HepG2 cells showed a slight change in cell morphology with a very rough periphery. Effervescence, crumb-like structures, and nuclear fragmentation occurred, apoptotic bodies appeared, the permeability of cell membrane increased, and a window for both stained AO and EB was observed, which is a typical apoptosis characteristic (Figure 3). We believe that the apoptotic bodies visualized in our results are a specific effect caused by the activation of the cell apoptosis process within cancerous cells by treatment with either of the compounds and not just a mere overall toxic effect of the chemicals.

Antifungal activity

The antifungal test of the compounds from *paris polyphylla* var. *yunnanensis* gave the results listed in Table 2. The results presented as minimal inhibitory concentration (MIC, mg.mL⁻¹) show that compounds 2, 3 and 4 convey relatively higher antifungal activities against *saccharomyces cerevisiae hansen* and *candida albicans* than compound 1. The MIC of compounds 2, 3 and 4 against *saccharomyces cerevisiae hansen* are 2.5 mg.mL⁻¹, 0.6 mg.mL⁻¹ and 0.6 mg.mL⁻¹, respectively. The MIC of compounds 2, 3 and 4 against *Candida albicans* are 1.2 mg.mL⁻¹, 0.6 mg.mL⁻¹ and 1.2 mg.mL⁻¹, respectively.

Structure-activity relationships

Compounds 2, 3 and 4, with the same steroidal saponin moiety, have different anticancer and antifungal activities. Compound 3 exhibits the strongest activity and also has the most sugar moieties among the three. This

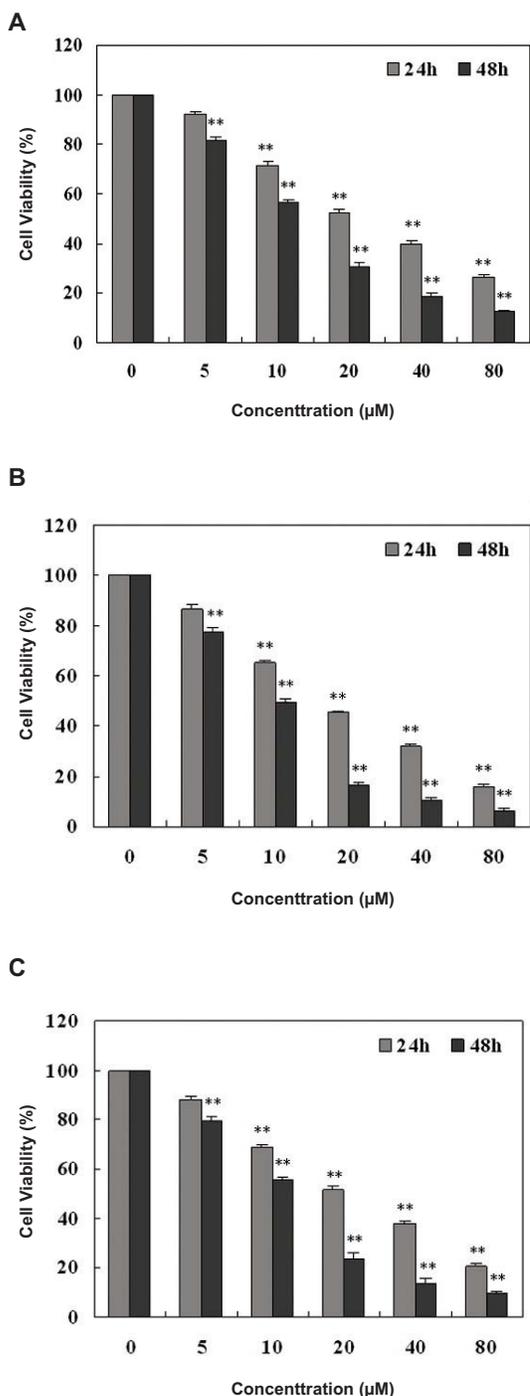


Figure 2. The effects of Compound 2, 3 and 4 on cell viability in HepG2 cells. (A) Compound 2; (B) Compound 3; and (C) Compound 4. The reported values are the means \pm SD (n=5). **p<0.01 represents a significant difference between the experimental and control value.

may suggest that there is a relation between the anticancer and antifungal activities of these compounds and the number of sugar moieties they contain. The water-solubility of these compounds increases in correlation with an increase in their sugar moiety number. Their anticancer and antifungal activity decreases in the order: compound 3 (4 sugar moieties) > compound 4 (3 sugar moieties) > compound 2 (2 sugar moieties). The hetero-sugar moiety causes the hetero-polarity of these compounds leading to different membrane permeability and selectivity, involved in the bioactivity of the compounds. The bioactivity-structure relationship of the compounds containing more sugar moieties than four remains unclear. Too many sugar moieties may lead to an excessively large polarity, which would then impair the solubility of the active ingredient within the lipophilic medium of the biomembrane and thus block its transmembrane transfer. Sugar moiety has been reported to play a critical role in the activity of steroid moiety. Diosgenin, stigmasterol and solanidine, which have the same skeleton and different sugar moieties, show hetero-activity in the cell cycle arrest (10). OSW-1, characterized as a cholestane disaccharide, is an extraordinarily potent antitumor saponin. Whereas, the glycon OSW-1 and the disaccharide OSW-1 show no significant inhibition to the growth and proliferation of tumor cells (11). These facts all allude to the importance of a precise number of sugar moieties in the bioactivity of saponins within living cells.

Conclusion

In this work, β -ecdysterone (compound 1) and three other compounds with the same steroidal saponin moiety were isolated from *Paris polyphylla var. yunnanensis* and their structures were identified. We found that compounds 2, 3 and 4 not only inhibited cancer cell proliferation but in addition induced cancer cell apoptosis. These three pennogenin steroidal saponins may provide clues for designing a range of novel semi-synthetic and synthetic compounds as medicinal anti-cancer agents in the near future.

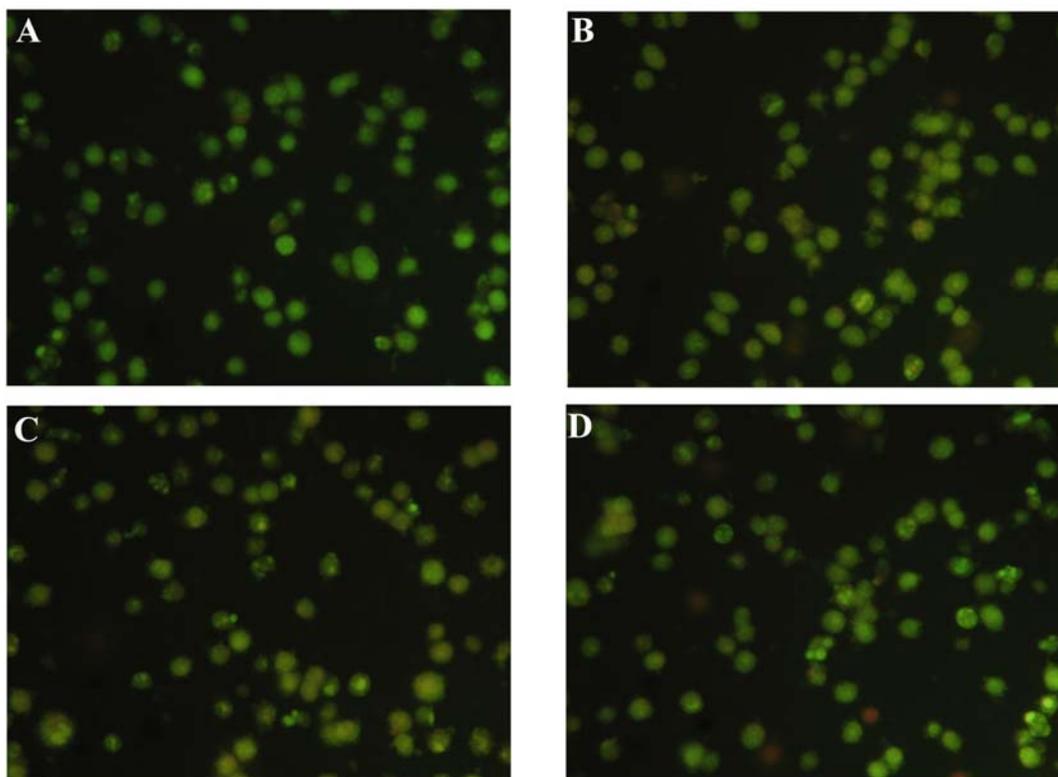


Figure 3. Apoptosis of HepG2 cells induced by compounds 2, 3 and 4 following 48 h treatment. After treatment with compounds 2, 3 and 4 for 48 h, the cells were stained with AO/EB at room temperature. The samples were observed under a fluorescence microscope immediately ($\times 100$). (A) Control; (B) 20 μ M Compound 2; (C) 20 μ M Compound 3; and (D) 20 μ M Compound 4.

Acknowledgments

The authors wish to thank the Fundamental Research Funds for the Central Universities (No. CDJRC10230002, CDJZR10230001) the National Natural Science Foundation of China (No. 20901086), the China Postdoctoral Science Foundation funded project (No. 20090450788) for their financial support.

References

- (1) Bureau of Chinese Herb Administration. *Chinese Herbal Medicine*. Shanghai Science and Technology Publishing Co., Shanghai (1999) 130.
- (2) Cheung JYN, Ong RCY, Suen YK, Ooi V, Wong HNC, Mak TCW, Fung KP, Yu B and Kong SK. Polyphyllin D is a potent apoptosis inducer in drug-resistant HepG2 cell. *Cancer Lett.* (2005) 217: 203-211.
- (3) Lee MS, Yuet-Wa JC, Kong SK, Yu B, Eng-Choon VO, Nai-Ching HW, Chung-Wai TM and Fung KP. Effects of polyphyllin D, a steroidal saponin in *Paris polyphylla*, in growth inhibition of human breast cancer cells and in xenograft. *Cancer Biol. Ther.* (2005) 4: 1248-1254.
- (4) Zhang XF, Cui Y, Huang JJ, Zhang YZ, Nie Z, Wang LF, Yan B Z, Tang YL and Liu Y. Immuno-stimulating properties of diosgenyl saponins isolated from *Paris polyphylla*. *Bioorg. Med. Chem. Lett.* (2007) 17: 2408-2413.
- (5) Mosmann T. Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. *J. Immunol. Methods.* (1983) 65: 55-63.
- (6) Li HB, Hu J, Chen JC and Qiu MH. Chemical Constituents of *Tinospora craveniana*. *Nat. Prod. Res. Develop.* (2005) 17: 125-127.
- (7) Huang Y, Cui LJ, Wang Q and Ye WC. Separation and identification of active constituents of *Paris vietnamensis*. *Acta Pharm. Sinica* (2006) 41: 361-364.
- (8) Matsuda H, Pongpiriyadacha Y, Morikawa T, Kishi A, Kataoka S and Yoshikawa M. Protective effects of steroid saponins from *Paris polyphylla* var. *yunnanensis* on ethanol-or indomethacin-induced gastric mucosal lesions in rats: structural requirement for activity and mode of action. *Bioorg. Med. Chem. Lett.* (2003) 13:

- 1101-1106.
- (9) Chen CX and Zhou J. The steroidal saponins from *Paris axialis*. *Acta Bot. Yunnan.* (1987) 9: 239-245.
- (10) Trouillas P, Corbière C, Liagre B, Duroux JL and Beneytout JL. Structure-function relationship for saponin effects on cell cycle arrest and apoptosis in the human 1547 osteosarcoma cells: a molecular modelling approach of natural molecules structurally close to diosgenin. *Bioorg. Med. Chem.* (2005) 13: 1141-1149.
- (11) Matsuya Y, Masuda S, Ohsawa N, Adam S, Tschamber T, Eustache J, Kamoshita K, Sukenaga Y and Nemoto H. Synthesis and antitumor activity of the estrane analogue of OSW-1. *Euro. J. Org. Chem.* (2005) 5: 803-808.

This article is available online at <http://www.ijpr.ir>
