

Isolation and Identification of Secondary Metabolites from the Aerial Parts of *Stachys lavandulifolia* Vahl.

Seyed Ebrahim Sajjadi^a, Seyed Mustafa Ghanadian^{a*}, Mohammad Rabbani^b and Fateme Tahmasbi^c

^aDepartment of Pharmacognosy, School of Pharmacy and Pharmaceutical Sciences, Isfahan University of Medical Sciences, Isfahan, Iran. ^bDepartment of Pharmacology and Toxicology, School of Pharmacy and Pharmaceutical Sciences, Isfahan University of Medical Sciences, Isfahan, Iran. ^cIsfahan Pharmaceutical Sciences Research Center, School of Pharmacy and Pharmaceutical Sciences, Isfahan University of Medical Sciences, Isfahan, Iran.

Abstract

Stachys lavandulifolia Vahl is an herbaceous wild plant native to Iran which is traditionally used in Iranian folk medicine as a mild sedative tea for reducing anxiety and for treatment of gastrointestinal disorders. Our previous study on ethyl acetate extract of *S. lavandulifolia* proved anti-anxiolytic activity and so the present study was designed to determine chemical components of this biologically active fraction. The extract was prepared using maceration method. Column chromatography and medium pressure liquid chromatography (MPLC) was used respectively to separate the fractions. Finally, some evaluated fractions were used for high pressure liquid (HPLC) and peak shaving recycle technique to achieve more purification. Separated compounds were determined using NMR analysis and mass spectroscopy. Six compounds have been isolated from ethylacetate extract of aerial parts of *S. lavandulifolia* including four flavonoids (apigenin, kumatakenin, penduletin and 4', 7-dihydroxy- 3, 5, 6-trimethoxy flavone), a labdan diterpenoid (labda-13-en-8, 15-diol), and an iridoid.

Keywords: *Stachys lavandulifolia*, labdan diterpenoid, flavonoid, monoterpen lacton, Lamiaceae

Introduction

The genus *Stachys* is one of the largest genera in the flowering plant family of Lamiaceae with about three hundred species distributed in Europe, Asia, Africa, Australasia, and North America(1).

Stachys lavandulifolia Vahl is an herbaceous wild plant native to Iran (2) which is used in Iranian folk medicine as a mild sedative tea for reducing anxiety and for treatment of gastrointestinal disorders (3).

Previous study on hydroalcoholic, polyphenolic, and boiled extracts of *S. lavandifolia* demonstrated their analgesic effect on animal models (4). In another study, antimicrobial and antioxidant activities of the essential oil of *S. lavandulifolia* were observed (5). Our previous study on *S. lavandulifolia* hydroalcoholic extract by the elevated plus-maze (EPM) model of anxiety in mice showed anxiolytic effect at therapeutically acceptable doses (6). Further study on petroleum ether, ethyl acetate, butanol and aqueous fractions on spontaneous motor activity and elevated

* Corresponding author:

E-mail: ghannadian@gmail.com

plus-maze behavior in mice showed interesting anxiolytic properties for ethyl acetate fraction (7) and thus, we decided to investigate the main chemical constituents responsible for anxiolytic effects of ethyl acetate extract of *S. lavandulifolia*.

In previous researches on polar fractions of methanol extract of this plant, lavandulifolioside A, lavandulifolioside B, verbascoside, leucosceptoside A, 5-O-allopyranosyloxy-aucubin together with three phenylethanoid glycosides were reported (8). The essential oil of aerial part of *S. lavandulifolia* was also analyzed by GC/MS method and germacrene-D (13.2%), β -phellandrene (12.7%), β -pinene (10.2%), myrcene (9.4%), α -pinene (8.4%) as well as Z- β -ocimene (5.8%) reported as the main components of the essential oil (9).

Experimental

General

NMR spectra were recorded on a Bruker Avance AV 400, using methanol-d₄ as solvent. ESI-MS spectra were measured in positive and negative mode on Shimadzu 2010EV LC-MS system (Shimadzu, Japan). Column chromatography runs were performed using Silica gel, 63-200 μ m (Merck) and polyamide SC6 (Macherey- Nagel, Duren, Germany). HPTLC was performed on silica gel GF₂₅₄ plates (Merck, Darmstadt, Germany). Plates were developed by Cerium sulphate (1 g in 5% H₂SO₄) or natural product reagent (1% methanolic diphenyl-boric acid-ethanolamine) and visualized by UV-fluorescent colours at 254 /366 nm UV lamps. Recycle HPLC was done on a modified Waters HPLC apparatus (Waters Assoc., Milford, MA, USA) at 250 nm using silica gel column (YMC-Pack SIL, 250 \times 20 mm, YMC Co., Kyoto, Japan).

Plant material

S. lavandulifolia was collected from Shahrekord city in Chaharmahal va Bakhtiari Province in the west of Iran. The plant was identified by the Department of Biology, Faculty of Science, University of Isfahan and also a voucher specimen (No. 1113) was deposited in the herbarium of the Isfahan Faculty of Pharmacy,

Isfahan University of Medical Sciences, Isfahan, Iran.

Extraction and isolation

Following with our previous study which ethyl acetate fraction of *S. lavandulifolia* showed anxiolytic effect based on spontaneous motor activity and elevated plus-maze behavior in Syrian mice (7), in the same order ethyl acetate extract of the plant (5 kg) was obtained. Filtration and vacuum evaporation resulted in a green mass (254 g). After partitioning between hexane and methanol in separatory funnel, lower defatted methanol fraction (56 g) was concentrated and column chromatographed on silica gel (63-200 μ m, 400 g) using hexane/acetone, with increasing polarity (5 \rightarrow 50 %) to afford five fractions (F1-F5). The fraction of F2 which was eluted by hexane: acetone (9:1) was applied on HPLC using chloroform: methanol (92:8) as solvent and yielded F2c2 (1) as a pure iridoid. F3 eluted by hexane: acetone (8:2) precipitated as yellowish crystals which was more purified by recycle HPLC using chloroform: methanol (90:10) to yield F3b1 (2) and F3b2 (4) as a labdane diterpene and a methoxylated flavonoid, respectively. Finally, F4 eluted with hexane: acetone (7:3) and F5 eluted with hexane: acetone (5:5) were applied on recycle HPLC using chloroform: methanol (85:15) and F4f2b (3), F5c2a (5), and F5e (6) with flavonoid structure were isolated.

Compound 1

Amorphous white powder, MW (g/mol): 196; yield: 0.00016%; ¹H-NMR (CDCl₃, 400 MHz): δ_{H} 5.71(s, 1H, H-10), 4.24 (m, 1H, H-5), 2.45 (1H, bd, J=14.1, H-4b), 1.93 (1H, bd, J=14.1, H-4a), 1.80 (s, 3H, H-7), 1.52 (1H, m, H-6b), 1.49 (s, 3H, H-1), 1.35(1H, m, H-6a), 1.30 (s, 3H, H-2). ¹³C-NMR data (CDCl₃, 100 MHz): 30.6 (C-1), 26.9 (C-2), 35.9 (C-3), 47.3 (C-4), 66.8 (C-5), 45.6 (C-6), 26.5 (C-7), 86.8 (C-8), 173.8 (C-9), 112.9 (C-10), 182.6 (C-11). EIMS *m/z* 196 [M], 178, 163, 152, 140, 125, 111, 95, 81, 69, 57.

Compound 2

Amorphous white powder, MW (g/mol): 308; yield: 0.00015%; ¹H-NMR (CDCl₃, 400 MHz):

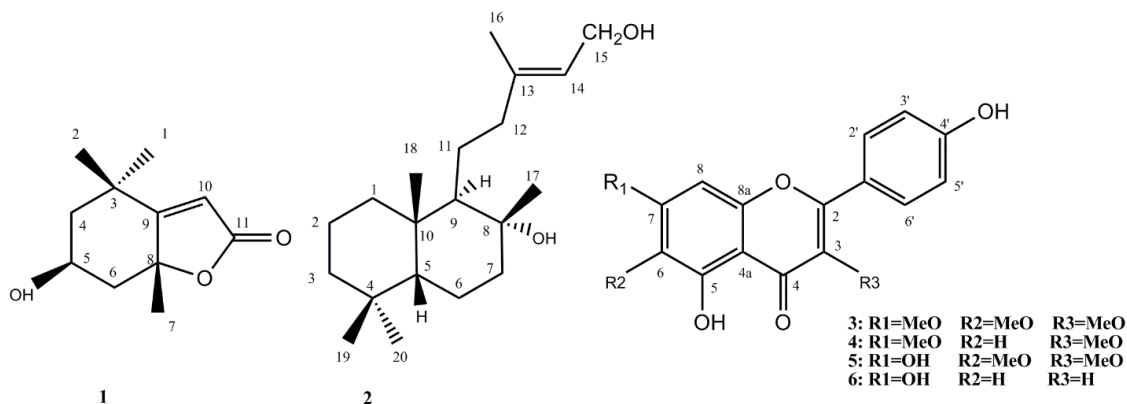


Figure 1. Chemical structures of compounds 1-6.

δ_{H} 5.38 (t, 1H, $J=6.8\text{Hz}$, H-14), 4.09 (d, 2H, $J=6.8\text{Hz}$, H-15), 1.70 (s, 3H, H-16), 1.14 (s, 3H, H-17), 0.90 (s, 3H, H-18), 0.86 (s, 3H, H-19), 0.84 (s, 3H, H-20). $^{13}\text{C-NMR}$ data (CDCl_3 , 100 MHz): 41.2 (C-1), 19.5 (C-2), 44.3 (C-3), 34.2 (C-4), 57.5 (C-5), 21.5 (C-6), 43.2 (C-7), 74.9 (C-8), 62.5 (C-9), 40.4 (C-10), 25.2 (C-11), 45.0 (C-12), 140.9 (C-13), 124.1 (C-14), 59.4 (C-15), 16.4 (C-16), 33.9 (C-17), 16.1 (C-18), 21.9 (C-19), 23.8 (C-20). Positive ESIMS m/z 331 $[\text{M}+\text{Na}]^+$, 309 $[\text{M}+\text{H}]^+$, 306, 290, 275.

Compound 3

Amorphous yellow powder, MW (g/mol): 344; yield: 0.0022%; $^1\text{H-NMR}$ (CDCl_3 , 400 MHz): δ_{H} 8.05 (d, 2H, $J=8.8\text{Hz}$, H-2', H-6'), 6.95 (d, 2H, $J=8.8\text{Hz}$, H-3', H-5'), 6.79 (s, 1H, H-8), 3.99 (s, 3H, OMe), 3.86 (s, 3H, OMe), 3.82 (s, 3H, OMe). $^{13}\text{C-NMR}$ data (CDCl_3 , 100 MHz): 158.6 (C-2), 139.5 (C-3), 180.3 (C-4), 107.3 (C-4a), 154.0 (C-5), 133.4 (C-6), 161.9 (C-7), 92.1 (C-8), 153.4 (C-8a), 122.5 (C-1'), 131.5 (C-2'), 116.6 (C-3'), 160.6 (C-4'), 116.6 (C-5'), 131.5 (C-6'), 60.6 (3-OMe), 61.1 (6-OMe), 57.0 (7-OMe). EIMS m/z 344 $[\text{M}]$, 329, 207, 197, 181, 167, 158, 149, 131, 121, 93, 69, 57.

Compound 4

Amorphous yellow powder, MW (g/mol): 314; yield: 0.001%; $^1\text{H-NMR}$ (CDCl_3 , 400 MHz): δ_{H} 8.05 (d, 2H, $J=8.8\text{Hz}$, H-2', H-6'), 6.95 (d, 2H, $J=9.2\text{Hz}$, H-3', H-5'), 6.66 (d, 1H, $J=2.4\text{Hz}$, H-8), 6.37 (d, 1H, $J=2.4\text{Hz}$, H-6), 3.91 (s,

3H, OMe), 3.81 (s, 3H, OMe). $^{13}\text{C-NMR}$ data (CDCl_3 , 100 MHz): 158.0 (C-2), 138.2 (C-3), 184.1 (C-4), 105.0 (C-4a), 154.3 (C-5), 98.8 (C-6), 161.4 (C-7), 93.0 (C-8), 154.3 (C-8a), 122.5 (C-1'), 131.2 (C-2'), 116.4 (C-3'), 158.2 (C-4'), 116.4 (C-5'), 131.2 (C-6'), 60.5 (3-OMe), 56.3 (7-OMe). Negative ESIMS m/z 313 $[\text{M}-1]^-$, 298, 269, 255, 227, 187, 166, 154, 136.

Compound 5

Amorphous yellow powder, MW (g/mol): 330; yield: 0.0035%; $^1\text{H-NMR}$ (CDCl_3 , 400 MHz): δ_{H} 7.89 (d, 2H, $J=8.8\text{Hz}$, H-2', H-6'), 6.83 (d, 2H, $J=8.8\text{Hz}$, H-3', H-5'), 6.41 (s, 1H, H-8), 3.78 (s, 3H, OMe), 3.67 (s, 3H, OMe). $^{13}\text{C-NMR}$ data (CDCl_3 , 100 MHz): 153.7 (C-2), 139.1 (C-3), 180.3 (C-4), 106.3 (C-4a), 153.8 (C-5), 132.6 (C-6), 158.8 (C-7), 95.0 (C-8), 158.2 (C-8a), 122.5 (C-1'), 131.4 (C-2'), 116.5 (C-3'), 161.8 (C-4'), 116.5 (C-5'), 131.4 (C-6'), 60.6 (3-OMe), 61.0 (6-OMe). Negative ESIMS m/z 329 $[\text{M}-1]^-$, 314, 299, 271, 227, 187, 166, 154, 125, 111.

Compound 6

Amorphous yellow powder, MW (g/mol): 269; yield: 0.0065%; $^1\text{H-NMR}$ (CDCl_3 , 400 MHz): δ_{H} 7.88 (s, 1H, OH), 7.73 (d, 2H, $J=8.8\text{Hz}$, H-2', H-6'), 6.85 (d, 2H, $J=8.8\text{Hz}$, H-3', H-5'), 6.49 (s, 1H, H-3), 6.37 (d, 1H, $J=2\text{Hz}$, H-8), 6.14 (d, 1H, $J=2\text{Hz}$, H-6). $^{13}\text{C-NMR}$ data (CDCl_3 , 100 MHz): 163.8 (C-2), 104.4 (C-3), 183.2 (C-4), 106.5 (C-4a), 163.2 (C-5), 100.5 (C-6), 166.4 (C-7), 95.3 (C-8), 159.0 (C-8a), 122.8 (C-1'),

129.4 (C-2'), 117.3 (C-3'), 163.3 (C-4'), 117.3 (C-5'), 122.8 (C-6'). Negative ESIMS m/z 269 [M-1]⁻; 257, 227, 195, 181, 173, 155, 129, 110.

Results and discussion

The ¹³C NMR and ¹H-NMR spectra of compound 1 indicated signals of three singlet methyl groups, two methylene, an oximethine, two quaternary carbons which one of them was oxygenated, a tri-substituted olefin bond and a lactone carbonyl with EIMS m/z 196 which resembled with those of a C11 iridoid named loliolide (10). The iridoids and secoiridoids form a large group of plant constituents that are found usually. They are mostly 4, 7-dimethylcyclopentapyran. But in the case of loliolide (C11 iridoid) and its related structures like actinidiolide and aeginetolide their biosynthesis pathway is different from usual C-10 iridoids and secoiridoids. They are generated in variety of plants from photo-oxygenation and degradation of carotenoids (11-12).

Compound 2 was obtained as a white powder. ¹³C-NMR data in addition to positive ESIMS at m/z 331 [M+Na]⁺ corresponded to the formula C₂₀H₃₆O₂. ¹H NMR spectrum showed signals attributed to five singlet methyls δ_H 1.70, 1.14, 0.90, 0.86, and 0.84. Further signals in ¹H NMR spectrum at δ_H 5.38 (t, 1H, J=6.8Hz, H-14) and 4.09 (d, 2H, J=6.8Hz, H-15) indicated the presence of a tri-substituted olefin bond and an external oxymethylen group. The ¹³C-NMR and DEPT spectra demonstrated twenty carbons comprised of five methyls, eight aliphatic methylenes (one oxygenated), two aliphatic methines, two aliphatic quaternary carbons (one oxygenated), one olefinic methine, and one quaternary olefinic carbon. Three degrees of unsaturation, and one olefinic bond suggested two rings in the structure. Comparing these characteristic signals with compound 6 isolated from *Leonurus heterophyllus* by Hung and coworkers (14) and compound 8 isolated from *Leonurus heterophyllus* by Giang and coworkers (13) and using HMBC spectra determined structure of compound 2 as labda-13-ene-8, 15-diol.

Compound 3 was isolated as a yellowish powder with orange fluorescence reaction to

Natural Product reagent (Diphenylboric acid 2-aminoethyl ester 1%). Molecular formula was determined by NMR data and negative EIMS m/z 344 [M]⁻ as C₁₈H₁₆O₇. Eleven degrees of unsaturation, one carbonyl carbon, and seven olefin bands indicated three rings in the structure. The ¹H-NMR spectrum showed a singlet olefin proton at δ 6.79 (s, 1H) described to H-8. *Ortho*-coupled proton signals (each two protons) at δ 8.05 (d, 2H, J=8.8Hz) and 6.95 (d, 2H, J=8.8Hz) were corresponded to H-2',6' and H-3',5'. Three singlet methoxy signals at δ_H 3.99 (δ_C 61.1), 3.86 (δ_C 60.6), 3.82 (δ_C 57.0) were located on C-3, C-6 and C-7, respectively based on long range HMBC correlation and cleavage pattern of compound in EI-Mass by retro-Diels-Alder mechanism including A (m/z 197), B (m/z 149) and C (m/z 93) ions. All of these data defined the structure of 3 as 3, 6, 7-trimethoxy-4',5'-dihydroxyflavone named as penduletin which was in agreement with literature data (15). Figure 2 shows the fragmentation pattern of this compound by cleavage of ring C via retro-Diels-Alder mechanism.

Compound 4 assigned the molecular formula of C₁₇H₁₄O₆ based on negative pseudo-molecular ion [M - H]⁻ at m/z 313 and ¹³C-NMR (BB and DEPT). ¹H & ¹³C-NMR spectral data cleared that compound 4 and 3 have similar structures but differed in lacking methoxy group at C-6. Comparison of the spectral data with literature data determined the structure as 7,3-dimethoxy-4',5'-dihydroxyflavone named as Kumatakenin (16). Similarly, NMR spectral data cleared that compounds 5 and 3 resembled each other, except for the 7-O-hydroxyl instead of 7-O-methoxyl group which was confirmed by pseudo-molecular ESI Mass ion [M - H]⁻ at m/z 329. It was defined as 6, 3-dimethoxy-4',5'-dihydroxyflavone (17). Compound 6 was proved to be 4',5,7-trihydroxyflavone named as apigenin (17).

Many species of the genus *Stachys* have been investigated and different kinds of secondary metabolites have been isolated, mainly flavonoids, iridoids and terpenoids from the aerial parts and the roots (18). In this investigation, an iridoid, a labdane type diterpene, three methoxylated flavonol derivatives, and a flavone were isolated.

Previous phytochemical studies showed

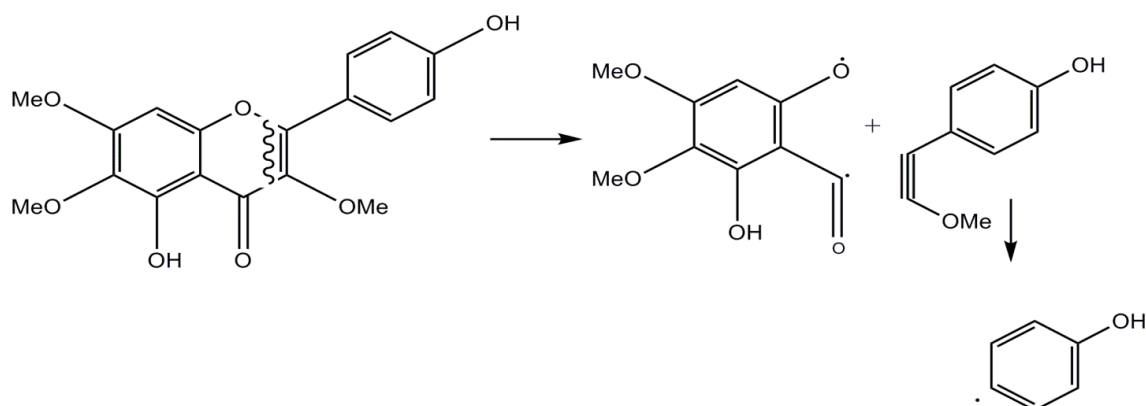


Figure 2. EI-Mass fragmentation pattern of compound 3. Cleavage of ring C by retro-Diels-Alder mechanism led to A (m/z 197), B (m/z 149), and C (m/z 93) ions.

presence of several types of flavonoids in *Stachys* genus. Some *Stachys* species are rich in flavonoids, for example 24 flavonoids were identified in *S. aegyptica* and 9 in *S. ionica* (19, 20). However in this study, three methoxylated flavonol derivatives, and a flavone were isolated from *S. lavandulifolia*. Compound 3 with trivial name of kumatakenin, previously was isolated from *Ballota hirsuta*, *Eupatorium illitum*, *Achillea kotschya*, and *Baccharis petiolata*. The flavonoids from *Eupatorium illitum* have been presented antiproliferative activity (22). Compound 4 with trivial name of penduletin, was previously isolated from *Achillea nobilis*, *Isocoma tenuisecta* and *Betula nigra* (21). Penduletin has showed antiproliferative property (23) and strong activity against enterovirus (24). Compound 5, was previously reported from *Achillea kotschya*, *Achillea nobilis*, and some other genera of Asteraceae and apigenin is also determined in many *Stachys* species (21).

The diterpenoids isolated from different species of *Stachys* are structurally different with labdane, kauran, clerodane, and abietan skeleton. Kauran diterpenoids are present in *S. silvatica*, *S. lanata*, and *S. sprumeri* and also *S. recta*, neo-clerodan derivatives are found in *S. annua*, *S. rosea*, and *S. recta*. An abietan derivative in *S. officinalise* and diterpenoids with labdane skeleton were also obtained from *S.*

mucronata, *S. plumasa*, and *S. menthifolia* (18, 25). The labdane diterpenoid which is isolated in this study was also previously reported from *S. menthifolia*, *Leonurus heterophyllus*, and *Cistus creticus* (11, 18).

References

- (1) Evans WC. *Trease and Evans' Pharmacognosy*. 13th ed, BailliereTindall, London (1989) 217.
- (2) Mozaffarian V. *A Dictionary of Iranian Plant Names*. Farhang Moaser, Tehran (1996) 552.
- (3) Amin GH. *Popular Medicinal Plants of Iran*. Vol. 1, Iranian Research Institute of Medicinal Plants, Tehran (1991) 47.
- (4) Hajhashemi V, Ghannadi A and Sedighifar S. Analgesic and anti-inflammatory properties of the hydroalcoholic, polyphenolic and boiled extracts of *Stachys lavandulifolia*. *Res. Pharm. Sci.* (2006) 2: 92-98.
- (5) Iscan G, Demirci B, Demirci F, Goger F, Kirimer N, Kose YB and Baser KHC. Antimicrobial and antioxidant activities of *Stachys lavandulifolia* subsp. *lavandulifolia* essential oil and its infusion. *Nat. Prod. Com.* (2012) 7: 1241-1244.
- (6) Rabbani M, Sajjadi SE and Zarei HR. Anxiolytic effects of *Stachys lavandulifolia* Vahl. on the elevated plus-maze model of anxiety in mice. *J. Ethnopharmacol.* (2003) 89: 271-276.
- (7) Rabbani M, Sajjadi SE and Jalali A. Hydroalcohol extract and fractions of *Stachys lavandulifolia* Vahl: effects on spontaneous motor activity and elevated plus maze behavior. *Phytother. Res.* (2005) 19: 854-858.
- (8) Delazar A, Delnavazi MR, Nahar L, Moghadam

- SB, Mojarab M, Gupta A, Williams AS, Mukhlesur Rahman M and Sarker SD. Lavandulifolioside B: a new phenylethanoid glycoside from the aerial parts of *Stachys lavandulifolia* Vahl. *Nat. Prod. Res.* (2011) 25: 8-16.
- (9) Javidnia K, Mojab F and Mojahedi SA. Chemical constituents of the essential oil of *Stachys lavandulifolia* Vahl from Iran. *Iran. J. Pharm. Res.* (2004) 3: 61-63.
- (10) Tanaka R and Matsunaga S. Loliolide and olean-12-en-3 β , 9 α , 11 α -triol from *Euphorbia supina*. *Phytochem.* (1989) 28: 1699-1702.
- (11) Isoe S, Hyeon SB, Katsumura Sand Sakan T. Photo-oxygenation of carotenoids. II. The absolute configuration of loliolide and dihydroactinidiolide. *Tetrahedron Lett.*, (1972) 13: 2517-2520.
- (12) Murai F, Tagawa M and Ohishi H. Absolute structure of kiwiionoside as a precursor of loliolide and actinidiolide, from *Actinidia chinensis*. *Planta Med.* (1992) 58: 112-113.
- (13) Hung TM, Luan TC, Vinh BT, Cuong TD and Min BS. Labdane-type diterpenoids from *Leonurus heterophyllus* and their cholinesterase inhibitory activity. *Phytother. Res.* (2011) 25: 611-4.
- (14) Giang PM, Son PT, Matsunami K and Otsuka H. New labdane-type diterpenoids from *Leonurus heterophyllus* Sw. *Chem. Pharm. Bull.* (2005) 53: 938-941.
- (15) Rodriguez E, Carman NJ, Vander Velde G, McReynolds JH, Mabry TJ, Irwin MA and Geissman TA. Methoxylated flavonoids from *Artemisia*. *Phytochem.* (1972) 11: 3509-14.
- (16) Calvert DJ, Cambie RC and Davis BR. ¹³C NMR spectra of polymethoxy- and methylenedioxy flavonols. *Org. Magn. Resonance* (1979) 12: 583-6.
- (17) Agrawal PK. *Carbon-13 NMR of Flavonoids*. Elsevier, Amsterdam (1989) Str. 2.233, 3.453. 132,160.
- (18) Tundis R, Peruzzi L and Menichini F. Phytochemical and biological studies of *Stachys* species in relation to chemotaxonomy: A review. *Phytochem.* (2014) 102: 7-39.
- (19) El-Ansari MA, Barron D, Abdalla MF, Saleh AM and Le Quere JL. Flavonoid constituents of *Stachys aegyptica*. *Phytochem.* (1991) 30: 1169-1173.
- (20) Meremeti A, Karioti A, Skaltsa H, Heilmann J and Sticher O. Secondary metabolites from *Stachys ionica*. *Biochem. Syst. Ecol.* (2004) 32: 139-151.
- (21) Harborne JB. *The Flavonoids, Advances in Research since 1980*. Chapman & Hall, London. (1994) 259-385, 441-479.
- (22) Castillo QA, Triana J, Eiroa JL, Padron JM, Plata GB, Abel-Santos EV, Báez LA, Rodríguez DC, Jiménez MA and Pérez-Pujols MF. Flavonoids from *Eupatorium illitum* and their antiproliferative activities. *Pharmacog. J.* (2015) 7: 179-181.
- (23) Moghaddam Gh, Ebrahimi SA, Rahbar-Roshandel N and Foroumadi A. Antiproliferative activity of flavonoids: influence of sesquimethoxylation state of the flavonoid structure. *Phytother. Res.* (2012) 26: 1023-1028.
- (24) Zho QC, Wang Y, Liu YP, Zhang RQ, Li X, Su WH, Long F, Luo XD and Peng T. Inhibition of enterovirus 71 replication by chrysosplentin and penduletin. *Eur. J. Pharm. Sci.* (2011) 44: 392-398.
- (25) Piozzi F and Bruno M. Diterpenoids from roots and aerial parts of the genus *Stachys*. *Rec. Nat. Prod.* (2011) 5: 1-11.