

Design, Synthesis and Biological Evaluation of New 1, 4-Dihydropyridine (DHP) Derivatives as Selective Cyclooxygenase-2 Inhibitors

Iman Sabakhi^{a,b}, Vigen Topuzyan^b, Zahra Hajimahdi^a, Bahram Daraei^c,
Hadi Arefi^a and Afshin Zarghi^{a*}

^aDepartment of Medicinal Chemistry, School of Pharmacy, Shahid Beheshti University of Medical Sciences, Tehran, Iran. ^bThe Scientific Thechnological Centre of Organic and Pharmaceutical Chemistry NASRAAL. Mnjoyan Institute of Fine Organic Chemistry, Yerevan, Armenia. ^cDepartment of Toxicology, Faculty of Medical Sciences, Tarbiat Modares University, Tehran, Iran.

Abstract

As a continuous research for discovery of new COX-2 inhibitors, chemical synthesis, in vitro biological activity and molecular docking study of a new group of 1, 4-dihydropyridine (DHP) derivatives were presented. Novel synthesized compounds possessing a COX-2 SO₂Me pharmacophore at the *para* position of C-4 phenyl ring, different hydrophobic groups (R₁) at C-2 position and alkoxy carbonyl groups (COOR₂) at C-3 position of 1, 4-dihydropyridine, displayed selective inhibitory activity against COX-2 isozyme. Among them, compound 5e was identified as the most potent and selective COX-2 inhibitor with IC₅₀ value of 0.30 μM and COX-2 selectivity index of 92. Molecular docking study was performed to determine probable binding models of compound 5e. The study showed that the *p*-SO₂Me-phenyl fragment of 5e inserted inside secondary COX-2 binding site (Arg⁵¹³, Phe⁵¹⁸, Gly⁵¹⁹, and His⁹⁰). The structure-activity relationships acquired reveal that compound 5e with methyl and ethoxycarbonyl as R₁ and COOR₂ substitutions has the necessary geometry to provide selective inhibition of the COX-2 isozyme and it can be a good basis for the development of new hits.

Keywords: Synthesis; 1, 4-Dihydropyridine (DHP) Derivatives; COX-2 Inhibitors; Molecular modeling.

Introduction

Cyclooxygenase (COX) also known as prostaglandin synthase (PGH) is a potent mediator of inflammation. Non-steroidal anti-inflammatory drugs (NSAIDs) bind to cyclooxygenase, thereby inhibiting the production of prostaglandins. However, inhibition of COXs may lead to undesirable side effects. Nowadays, it is well established that

there are at least two COX isozymes, COX-1 and COX-2 (1). The constitutive COX-1 isozyme is produced in a variety of tissues and appears to be important to the maintenance of physiological functions such as gastric protection and vascular homeostasis (2, 3). As COX-2 is usually specific to inflamed tissue, there is much less gastric irritation associated with COX-2 inhibition. This has led to intense efforts in searching for potent and selective COX-2 inhibitors which could provide anti-inflammatory drugs with fewer risks. Several classes of compounds having selective COX-2 inhibitory activity have been reported in

* Corresponding author:

E-mail: zarghi@sbmu.ac.ir

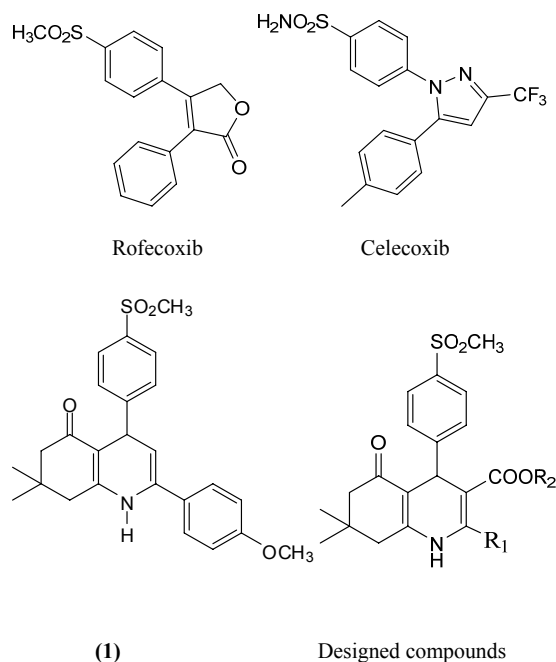


Figure 1. Selective COX-2 inhibitors (Rofecoxib, Celecoxib), lead compound (1) and designed scaffold.

the literature such as rofecoxib and celecoxib (Figure 1). Selective cyclooxygenase-2 (COX-2) inhibitors frequently belong to a class of diaryl heterocycles that possess two vicinal rings attached to a central heterocyclic scaffold in conjunction with a COX-2 pharmacophore such as a *para*-SO₂Me substituent on one of the rings (4-6). As an initial attempt to discover novel COX-2 inhibitor with selectivity and safety profile, we have recently reported several investigations describing the design, synthesis, and a molecular modeling study for a group of 5-oxo-1,4,5,6,7,8-hexahydroquinoline regioisomers including compound (1) from our compound library showed a good COX-2 inhibitory activity (Figure 1) (7). In continuation of our ongoing research work directed towards the development of selective COX-2 inhibitors, we have focused on the modification of compound (1) and designed some novel 1,4-dihydropyridines possessing *p*-SO₂Me-phenyl moiety at C-4 position, different hydrophobic groups at C-2 position (R₁) and different alkoxy carbonyl (COOR₂) groups at the C-3 position (Figure 1). 1,4-Dihydropyridines (DHP) are biologically

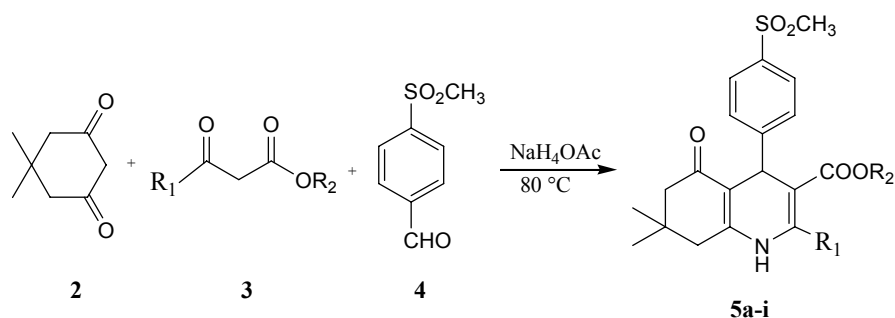
and synthetically important class of compounds in the field of drugs and pharmaceuticals and have attracted attention of synthetic chemists due to their pharmacological properties (8, 9). The Hantzsch reaction is a well-known method for synthesizing of dihydropyridines (10). Hantzsch reaction is a kind of multi component reactions (MCRs) which have gained wide applicability in the field of synthetic organic chemistry as they increase the efficiency of the reaction and decrease the number of laboratory operations along with quantities of solvent and chemicals (11, 12).

In this study novel 1, 4-dihydropyridine derivatives were prepared according to Hantzsch reaction and evaluated for in vitro COX-1/COX-2 isozyme inhibition. We also performed docking studies to determine the orientation of the synthesized compounds in the COX-2 active site which led to the better understanding of the structure-activity relationship in designed COX-2 inhibitors.

Experimental

General

All chemicals and solvents used in this study were purchased from Merck AG and Aldrich Chemical. Melting points were determined using a Thomas-Hoover capillary apparatus. Infrared spectra were acquired using a Perkin Elmer Model 550 SE spectrometer. A Bruker AM-300 NMR spectrometer was used to acquire ¹H NMR spectra with TMS as internal standard. Coupling constant (*J*) values are estimated in hertz (Hz) and spin multiples are given as s (singlet), d (double), t (triplet), q (quartet), m (multiplet), and br (broad). Low-resolution mass spectra were acquired with an MAT CH5/DF (Finnigan) mass spectrometer that was coupled on line to a Data General DS 50 data system. Electron-impact ionization was performed at an ionizing energy of 70 eV with a source temperature of 250°C. Elemental microanalyses, determined for C and H, were within ±0.4% of theoretical values. All chemicals and solvents used in this study were purchased from Merck AG and Aldrich Chemical. Melting points were determined with a Thomas-Hoover capillary apparatus. Infrared spectra were acquired using a Perkin Elmer Model 1420



Scheme 1. Synthesis of 1, 4-dihydropyridine derivatives (5a-i).

spectrometer. A Bruker FT-500 MHz instrument (Bruker Biosciences, USA) was used to acquire ¹H NMR spectra with TMS as internal standard. Chloroform-D was used as solvents. Coupling constant (*J*) values are estimated in hertz (Hz) and spin multiples are given as s (singlet), d (double), t (triplet), q (quartet), m (multiplet) and br (broad). The mass spectral measurements were performed on a 6410 Agilent LCMS triple quadrupole mass spectrometer (LCMS) with an electrospray ionization (ESI) interface.

Chemistry

Preparation of 1, 4-dihydropyridine derivatives based on Hantzsch method is shown in Scheme 1. Accordingly, a mixture of 5, 5-dimethyl-1,3-cyclohexandione (2), appropriate β-oxoesters (3) and 4-(methylsulfonyl)benzaldehyde (4) in the presence of ammonium acetate was refluxed in methanol to obtain target compounds (5a-i) in 54-95% yield. The structure of the synthesized compounds was confirmed by IR, ¹H NMR and ESI-MS.

General procedure for the synthesis of 1, 4-dihydropyridine derivatives (5a-i)

A mixture of β-oxoesters (1 mmol), 4, 4-(5, 5)-dimethyl-1,3-cyclohexandione (1 mmol) and 4-(methylsulfonyl)benzaldehyde (1 mmol) in the presence of ammonium acetate (4 mmol) was refluxed in methanol at 80 °C for overnight. After completion of the reaction, the mixture was cooled to room temperature; ethanol (10 mL) was added to dilute mixture. The mixture was poured into 80 mL ice water the precipitate was filtered off and washed with water. The

crude products were purified by recrystallization from ethanol to give final products.

Methyl-1, 4, 5, 6, 7, 8-hexahydro-2, 7, 7-trimethyl-4-(4-(methylsulfonyl)phenyl)-5-oxoquinoline-3-carboxylate (5a)

Yield, 78%; mp: 244-245 °C; IR (KBr disk) ν (cm⁻¹): 1150, 1300 (SO₂); 1400-1600 (aromatic); 1689 (C=O); 3356 (NH); ¹H NMR (CDCl₃, 500 MHz): δ 0.91 (s, 3H, CH₃), 1.12 (s, 3H, CH₃), 2.16-2.20 (m, 2H, dihydroquinoline H₈), 2.26-2.29 (m, 2H, dihydroquinoline H₆, *J*=15.8 Hz), 2.45 (s, 3H, CH₃), 3.04 (s, 3H, SO₂Me), 3.64 (s, 3H, CO₂CH₃), 5.17 (s, 1H, dihydroquinoline H₄), 5.84 (s, 1H, NH), 7.54 (d, 2H, methanesulfonyl phenyl H₂ & H₆, *J*=7.6 Hz), 7.81 (d, 2H, methanesulfonyl phenyl H₃ & H₅, *J*=7.6 Hz); LC-MS (ESI) *m/z*: 404.3 (M+1, 100); Anal. Calcd. for C₂₁H₂₅NO₅S: C, 62.51; H, 6.25; N, 3.47. Found: C, 62.81; H, 6.45; N, 3.59.

Methyl-2-amino-1, 4, 5, 6, 7, 8-hexahydro-7, 7-dimethyl-4-(4-(methylsulfonyl)phenyl)-5-oxoquinoline-3-carboxylate (5b)

Yield, 64%; mp: 177-179 °C; IR (KBr disk) ν (cm⁻¹): 1150, 1300 (SO₂); 1400-1600 (aromatic); 1697 (C=O); 3350 (NH); ¹H NMR (CDCl₃, 500 MHz): δ 0.99 (s, 3H, CH₃), 1.12 (s, 3H, CH₃), 2.20 (d, 1H, dihydroquinoline H₈, *J*=16.3 Hz), 2.29 (d, 1H, dihydroquinoline H₈, *J*=16.3 Hz), 2.48 (s, 2H, dihydroquinoline H₆), 3.07 (s, 3H, SO₂Me), 3.62 (s, 3H, CO₂CH₃), 4.81 (s, 1H, NH), 6.27 (br s, 2H, NH₂), 7.49 (d, 2H, methanesulfonyl phenyl H₂ & H₆, *J*=8.2 Hz), 7.86 (d, 2H, methanesulfonyl phenyl H₃ & H₅, *J*=8.2 Hz); LC-MS (ESI) *m/z*: 405.1 (M+1,

100); Anal. Calcd. for $C_{20}H_{24}N_2O_5S$: C, 59.39; H, 5.98; N, 6.93. Found: C, 59.71; H, 6.25; N, 6.59.

Methyl-2-ethyl-1, 4, 5, 6, 7, 8-hexahydro-7,7-dimethyl-4-(4-(methylsulfonyl)phenyl)-5-oxoquinoline-3-carboxylate (5c)

Yield, 87%; mp: 186.5-188°C; IR (KBr disk) ν (cm^{-1}): 1150, 1300 (SO_2); 1400-1600 (aromatic); 1697 (C = O); 3369 (NH); 1H NMR ($CDCl_3$, 500 MHz): δ 0.94 (s, 3H, CH_3), 1.07 (s, 3H, CH_3), 1.36 (t, 3H, CH_3), 2.17-2.44 (m, 4H, dihydroquinoline H_6 & H_8), 2.85 (q, 2H, CH_2), 3.04 (s, 3H, SO_2Me), 3.64 (s, 3H, CO_2CH_3), 5.19 (s, 1H, dihydroquinoline H_4), 5.80 (br s, 1H, NH), 7.51 (d, 2H, methanesulfonyl phenyl H_2 & H_6 , $J=8.1$ Hz), 7.81 (d, 2H, methanesulfonyl phenyl H_3 & H_5 , $J=8.2$ Hz); LC-MS (ESI) m/z : 418.4 (M+1, 100); Anal. Calcd. for $C_{22}H_{27}NO_5S$: C, 63.29; H, 6.52; N, 3.35. Found: C, 62.91; H, 6.35; N, 3.50.

Methyl-1, 4, 5, 6, 7, 8-hexahydro-2-isopropyl-7, 7-dimethyl-4-(4-(methylsulfonyl)-5-oxoquinoline-3-carboxylate (5d)

Yield, 54%; mp: 163-164°C; IR (KBr disk) ν (cm^{-1}): 1150, 1300 (SO_2); 1400-1600 (aromatic); 1657 (C=O); 3352 (NH); 1H NMR ($CDCl_3$, 500 MHz): δ 0.92 (s, 3H, CH_3), 1.11 (s, 3H, CH_3), 1.21 (d, 3H, CH_3 , $J=6.9$ Hz), 1.27 (d, 3H, CH_3 , $J=7.0$ Hz), 2.19 (d, 1H, dihydroquinoline H_8 , $J=16.3$ Hz), 2.29 (m, 2H, dihydroquinoline H_8 & H_6 , $J=15.0$ Hz), 2.45 (d, 1H, dihydroquinoline H_6 , $J=16.6$ Hz), 3.04 (s, 3H, SO_2Me), 3.63 (s, 3H, CO_2CH_3), 4.27 (m, 1H, CH), 5.19 (s, 1H, dihydroquinoline H_4), 6.07 (s, 1H, NH), 7.50 (d, 2H, methanesulfonyl phenyl H_2 & H_6 , $J=8.3$ Hz), 7.80 (d, 2H, methanesulfonyl phenyl H_3 & H_5 , $J=8.3$ Hz); LC-MS (ESI) m/z : 432.2 (M+1, 100); Anal. Calcd. for $C_{23}H_{29}SO_5N$: C, 64.01; H, 6.77; N, 3.25. Found: C, 64.21; H, 6.95; N, 3.19.

Ethyl-1, 4, 5, 6, 7, 8-hexahydro-2, 7, 7-trimethyl-4-(4-(methylsulfonyl)phenyl)-5-oxoquinoline-3-carboxylate (5e)

Yield, 81%; mp: 180.7-182.3°C; IR (KBr disk) ν (cm^{-1}): 1150, 1300 (SO_2); 1400-1600 (aromatic); 1685 (C = O); 3359 (NH); 1H NMR ($CDCl_3$, 500 MHz): δ 0.91 (s, 3H, CH_3), 1.10 (s, 3H, CH_3), 1.24 (t, 3H, CH_3), 2.12-2.15 (d, 1H, dihydroquinoline H_8), 2.18-2.28 (m, 2H, dihydroquinoline H_8 &

H_6), 2.35-2.38 (d, 1H, dihydroquinoline H_6), 2.40 (s, 3H, CH_3), 3.01 (s, 3H, SO_2Me), 4.06 (m, 2H, CH_2), 5.14 (s, 1H, dihydroquinoline H_4), 7.09 (s, 1H, NH), 7.51 (d, 2H, methanesulfonyl phenyl H_2 & H_6 , $J=8.0$ Hz), 7.76 (d, 2H, methanesulfonyl phenyl H_3 & H_5 , $J=8.0$ Hz); LC-MS (ESI) m/z : 418.1 (M+1, 100); Anal. Calcd. for $C_{22}H_{27}NO_5S$: C, 63.29; H, 6.52; N, 3.35. Found: C, 63.41; H, 6.75; N, 3.42.

Ethyl-1, 4, 5, 6, 7, 8-hexahydro-7,7-dimethyl-4-(4-(methylsulfonyl)phenyl)-5-oxo-2-propylquinoline-3-carboxylate (5f)

Yield, 54%; mp: 163-164°C; IR (KBr disk) ν (cm^{-1}): 1150, 1300 (SO_2); 1400-1600 (aromatic); 1658 (C = O); 3305 (NH); 1H NMR ($CDCl_3$, 500 MHz): δ 0.88 (s, 3H, CH_3), 1.05 (t, 3H, CH_3), 1.12 (s, 3H, CH_3), 1.27 (t, 3H, CH_3), 1.71 (m, 4H, $2CH_2$), 2.16-2.20 (d, 1H, dihydroquinoline H_8 , $J=16.2$ Hz), 2.26-2.29 (m, 2H, dihydroquinoline H_6 & H_8), 2.39-2.42 (d, 1H, dihydroquinoline H_6 , $J=16.0$ Hz), 3.04 (s, 3H, SO_2Me), 4.07 (m, 2H, CH_2), 5.19 (s, 1H, dihydroquinoline H_4), 5.93 (br s, 1H, NH), 7.52 (d, 2H, methanesulfonyl phenyl H_2 & H_6 , $J=7.9$ Hz), 7.80 (d, 2H, methanesulfonyl phenyl H_3 & H_5 , $J=7.8$ Hz); LC-MS (ESI) m/z : 446.2 (M+1, 100); Anal. Calcd. for $C_{24}H_{31}NO_5S$: C, 64.69; H, 7.01; N, 3.14. Found: C, 63.89; H, 6.95; N, 3.32.

Ethyl-1, 4, 5, 6, 7, 8-hexahydro-7,7-dimethyl-4-(4-(methylsulfonyl)phenyl)-5-oxo-2-phenylquinoline-3-carboxylate (5g)

Yield, 87%; mp: 187.9-189 °C; IR (KBr disk) ν (cm^{-1}): 1150, 1300 (SO_2); 1400-1600 (aromatic); 1687 (C = O); 3344 (NH); 1H NMR ($CDCl_3$, 500 MHz): δ 0.89 (t, 3H, CH_3), 0.97 (s, 3H, CH_3), 1.14 (s, 3H, CH_3), 2.19-2.32 (m, 3H, dihydroquinoline H_6 & H_8), 2.45-2.48 (d, 1H, dihydroquinoline H_6 , $J=16.6$ Hz), 3.03 (s, 3H, SO_2Me), 3.86 (m, 2H, CH_2), 5.28 (s, 1H, dihydroquinoline H_4), 6.07 (s, 1H, NH), 7.36 (m, 2H, benzyl H_3 & H_4), 7.45 (m, 3H, benzyl H_2 , H_5 & H_6), 7.68 (d, 2H, methanesulfonyl phenyl H_2 & H_6 , $J=7.6$ Hz), 7.85 (d, 2H, methanesulfonyl phenyl H_3 & H_5 , $J=7.5$ Hz); LC-MS (ESI) m/z : 480.2 (M+1, 100); Anal. Calcd. for $C_{27}H_{29}NO_5S$: C, 67.62; H, 6.09; N, 2.92. Found: C, 63.96; H, 6.25; N, 3.12.

t-Butyl-1, 4, 5, 6, 7, 8-hexahydro-2,7,7-trimethyl-4-(4-methanesulfonyl-phenyl)-5-oxoquinoline-3-carboxylate (5h)

Yield, 95%; mp: 163-164°C; IR (KBr disk) ν (cm⁻¹): 1150, 1300 (SO₂); 1400-1600 (aromatic); 1694 (C = O); 3300-3500 (NH); ¹H NMR (CDCl₃, 500 MHz): δ 0.95 (s, 3H, °C H₃), 1.11 (s, 3H, CH₃), 1.37 (s, 9H, CH₃), 2.14 (d, 1H, dihydroquinoline H₈, *J*=16.3 Hz), 2.26 (d, 1H, dihydroquinoline H₈, *J*=15.9 Hz), 2.36-2.39 (d, 2H, dihydroquinoline H₆), 2.41 (s, 3H, CH₃), 3.03 (s, 3H, SO₂Me), 5.10 (s, 1H, dihydroquinoline H₄), 5.91 (br s, 1H, NH), 7.54 (d, 2H, methanesulfonyl phenyl H₂ & H₆, *J*=8.2 Hz), 7.81 (d, 2H, methanesulfonyl phenyl H₃ & H₅, *J*=8.2 Hz); LC-MS (ESI) *m/z*: 446.2 (M+1, 100); Anal. Calcd. for C₂₄H₃₁NO₅S: C, 64.69; H, 7.01; N, 3.14. Found: C, 64.89; H, 7.21; N, 3.22.

Benzyl-1, 4, 5, 6, 7, 8-hexahydro-2, 7, 7-trimethyl-4-(4-(methylsulfonyl)phenyl)quinoline-3-carboxylate (5i)

Yield, 87%; mp: 136.9-138.9 °C; IR (KBr disk) ν (cm⁻¹): 1150, 1300 (SO₂); 1400-1600 (aromatic); 1694 (C=O); 3557 (NH); ¹H NMR (CDCl₃, 500 MHz): δ 0.86 (s, 3H, CH₃), 1.04 (s, 3H, CH₃), 2.01-2.07 (d, 1H, dihydroquinoline H₈, *J*=16.3 Hz), 2.17-2.20 (d, 1H, dihydroquinoline H₈, *J*=16.4 Hz), 2.20-2.36 (q, 2H, dihydroquinoline H₆), 2.38 (s, 3H, CH₃), 2.96 (s, 3H, SO₂Me), 4.98 (s, 2H, CH₂), 5.10 (s, 1H, dihydroquinoline H₄), 7.11-7.12 (m, 2H, benzyl H₂ & H₆), 7.26 (m, 3H, benzyl H₃, H₄ & H₅), 7.41 (d, 2H, methanesulfonyl phenyl H₂ & H₆, *J*=8.3 Hz), 7.67 (d, 2H, methanesulfonyl phenyl H₃ & H₅, *J*=8.3 Hz); LC-MS (ESI) *m/z*: 480.2 (M+1, 100); Anal. Calcd. for C₂₇H₂₉NO₅S: C, 67.62; H, 6.09; N, 2.92. Found: C, 67.32; H, 5.84; N, 3.02.

Molecular Modeling

The active compound was selected for docking studies which performed using Autodock software Version 4.0. The ligand molecule was constructed using the Chem Draw and was energy minimized for 1000 iterations reaching a convergence of 0.01 kcal/mol Å. The coordinates of the X-ray crystal structure of COX-2 enzyme was obtained from the RCSB

Protein Data Bank (3NT1) and the protein structure was prepared for docking. First of all, co-crystallized ligand and all water molecules were removed from crystal protein. Polar hydrogens were added and non polar hydrogens were merged, finally Kollman united atom charge and atom type parameter was added to 3NT1. Grid map dimensions (20×20×20) were set surrounding active site. Lamarckian genetic search algorithm was employed and docking run was set to 50. The aim of docking is to search for suitable binding configuration between the ligands and the rigid protein. These docked structures were very similar to the minimized structures provided initially. The quality of the docked structures was determined by measuring the intermolecular energy of the ligand-enzyme assembly (13).

Result and Discussion

A group of 1,4-dihydropyridine derivatives possessing a MeSO₂ at the *para*-position of the C-4 phenyl ring, alkyl groups (R₁) at the C-2 position and alkyloxycarbonyl groups (COOR₂) at the C-3 position were prepared and evaluated for their ability to inhibit COX-1 and COX-2 using chemiluminescent kit (Cayman chemical, MI, USA) according to our previously reported method (14). Potent and selective COX-2 inhibitor, celecoxib was used as a reference compound in the COX activity assay. All experiments were carried out at least three times and the data of inhibitory effects were summarized in Table 1.

As shown in Table 1, all compounds except 5i and 5g (IC₅₀ > 100 μM) displayed moderate to good inhibitory activities against COX-2 and were more potent inhibitor of COX-2 (IC₅₀ = 0.3-1.38 μM range) than COX-1 (IC₅₀ = 22.9-46.1 μM range) with COX-2 selectivity indexes (SI) in the range of 18.2-92.0. However, in all cases, the measured activities were lower than that of celecoxib. Our results indicated that different hydrophobic substituents at C-2 and C-3 position of 1, 4-dihydropyridine core affected the activity of the target molecules. In compounds series possessing methoxycarbonyl as COOR₂ group (5a-d), replacement of methyl (5a, IC₅₀ = 0.48 μM) at C-2 position with other alkyl groups such

Table 1. *In-vitro* COX-1 and COX-2 enzyme inhibition data for compounds 5a-i.

Compound	R ₁	R ₂	IC ₅₀ (μ M) ^a		COX-2 S.I. ^b
			COX-1	COX-2	
5a	CH ₃	CH ₃	30.2	0.48	62.9
5b	NH ₂	CH ₃	31.0	0.44	70.4
5c	CH ₂ CH ₃	CH ₃	30.7	0.59	52.1
5d	CH(CH ₃) ₂	CH ₃	27.6	0.62	43.1
5e	CH ₃	CH ₂ CH ₃	27.6	0.30	92.0
5f	CH ₂ CH ₂ CH ₃	CH ₂ CH ₃	25.2	1.38	18.2
5g	C ₆ H ₅	CH ₂ CH ₃	46.1	>100	-
5h	CH ₃	C(CH ₃) ₃	25.5	0.40	63.7
5i	CH ₃	CH ₂ C ₆ H ₅	22.9	>100	-
celecoxib			24.3	0.06	405

^aValues are mean values of two determinations acquired using an ovine COX-1/COX-2 assay kit, where the deviation from the mean is < 10% of the mean value.

^b*In-vitro* COX-2 selectivity index (COX-1 IC₅₀/ COX-2 IC₅₀).

as ethyl (5c, IC₅₀ = 0.59 μ M) and isopropyl (5d, IC₅₀ = 0.62 μ M) slightly decreased the COX-2 inhibitory activity. Compound 5b showed approximately similar potency (IC₅₀ = 0.44 μ M) to compound 5a. This may be due to isosteric replacement of methyl group with NH₂ group in compound 5b. It is found that replacement of methoxycarbonyl with ethoxycarbonyl as R₂ group in compound 5a resulted in compound 5e with improved COX-2 inhibitory effect (IC₅₀ = 0.30 μ M). Introduction of larger groups such as propyl and phenyl at C-2 position of compound 5e led to compounds 5f and 5g with significant loss of activities. The experimental results showed that *t*-butoxycarbonyl as COOR₂ group is well tolerated and the corresponding compound, 5h exhibited IC₅₀ value of 0.40 M. In addition, modification of ethoxycarbonyl group to benzyloxycarbonyl group in compound 5e led to compound 5i with no activity (IC₅₀ > 100 μ M). This may be due to large size of substitution and resulting steric hindrance. The effects of substituents introduced into the 1, 4-dihydropyridines moiety of compounds demonstrated that methyl and ethoxycarbonyl groups were the most appropriate substitutions at C-2 and C-3 positions, respectively and the corresponding compound, 5e was the most potent COX-2 inhibitor in this series with IC₅₀ value of 0.30 μ M and COX-2 selectivity index of 92.

Molecular docking studies help to understand the various interactions between the most active ligand (5e) and enzyme active sites in details. According to docking studies results (Figure 2), it is clear that *p*-SO₂Me-phenyl moiety of compound 5e inserts deep inside the COX-2 active site pocket and forming hydrogen bond with Arg⁵¹³ (distance = 4.8 Å) and His⁹⁰ (distance = 3.1 Å). In addition, the N-H of the 1, 4-dihydropyridine scaffold interacts with C=O of Val³⁴⁹ (distance = 4.0 Å). Moreover, the carbonyl group of central ring and ethoxycarbonyl bind to Arg¹²⁰ (distance = 2.8 Å) and Gly⁵²⁶ (distance = 3.9 Å) through hydrogen bonds, respectively. Molecular docking studies associated with experimental results showed that compound 5e possesses the pharmacophoric requisites for COX-2 inhibition.

Conclusion

In conclusion, new 1, 4-dihydropyridine derivatives were synthesized and evaluated for COX-1/COX-2 inhibition. Among them, compound 5e exhibited good COX-2 inhibitory activity and selectivity (IC₅₀ = 0.30 μ M and COX-2 selectivity index = 92). Experimental results in conjunction with molecular docking studies indicated that compound 5e with methyl and ethoxycarbonyl groups as R₁ and COOR₂ substitutions could interact appropriately

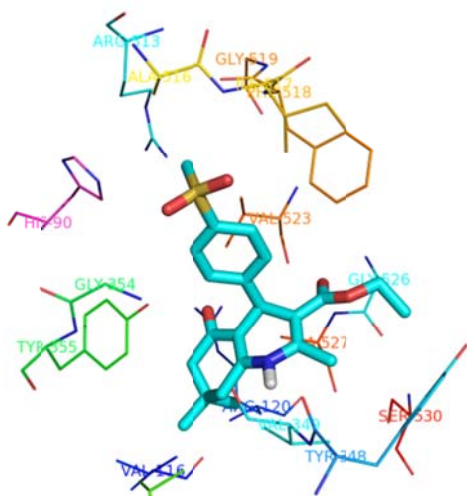


Figure 2. Binding mode of compound **5e** in the COX-2 active site.

with COX-2 active site. Therefore, this compound provides a promising lead for further development.

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