

## Free Radical Scavenging Principles of *Salvia reuterana* Boiss. Aerial Parts

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### Abstract

*Salvia reuterana* Boiss. is an aromatic perennial plant traditionally used for its anxiolytic and sedative properties. In the present study, various fractions and essential oil of *S. reuterana* aerial parts were investigated to find its free radical scavenging principles. Hydroalcoholic fraction with IC<sub>50</sub> value of 112.6 ± 3.2 µg mL<sup>-1</sup> in DPPH assay demonstrated the highest free radical scavenging activity and was selected to further phytochemical investigation. RP-18 and Sephadex LH-20 column chromatography of the hydroalcoholic fraction resulted in the isolation and structural elucidation of four phenolic derivatives, including apigenin-7-O-β-D-glucopyranoside (**1**), luteolin-7-O-β-D-glucopyranoside (**2**), rosmarinic acid (**3**), and luteolin (**4**). Isolated compounds showed potent free radical scavenging activities (5.1-34.2 µg mL<sup>-1</sup>), compared with BHT (21.30 ± 1.9 µg mL<sup>-1</sup>). Twenty four compounds were also identified in GC-MS analysis of the plant essential oil, of which benzyl benzoate (26.64%), *n*-hexyl benzoate (22.99%) and *n*-hexyl isovalerate (6.04%) were the main compounds. The results of the present study introduced *S. reuterana* as a valuable source of natural phenolic antioxidants which can be utilized in prevention of oxidative stress related diseases. Moreover, interesting composition of *S. reuterana* essential oil, dominated by non-terpenes compounds (76.17%) especially aromatic derivatives, make it an appropriate candidate for more detailed studies.

**Keywords:** *Salvia reuterana* Boiss; DPPH; Flavonoid; Rosmarinic acid; Essential oil; GC-MS; Benzyl benzoate.

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### Introduction

Nowadays, the role of free radicals, particularly reactive oxygen species (ROS), have been well recognized in development of many pathological disorders such as cardiovascular diseases, diabetes, cancers, inflammatory, and neurodegenerative diseases (e.g. Alzheimer's disease, Parkinson's disease, etc.) (1). So, in recent years plants have

received considerable attention as source of potent and safe natural free radical scavengers to prevent oxidative stress related diseases (2).

The genus *Salvia* L., commonly known as Sage, is one of the largest genera in the Lamiaceae family, mainly distributed in temperate and subtropical regions of the world (3). In the flora of Iran this genus is represented by 61 species, including *Salvia reuterana* Boiss. (4). This perennial aromatic plant is known as "Maryam Goli-e Esfahani" in Persian and its flowering aerial parts are traditionally used

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in some parts of Iran as anti-depressant, as well as for the treatment of gastrointestinal disorders, eye pains and colds (5, 6). Previous pharmacological studies have confirmed the anxiolytic (7), hypnotic (8), antidiabetic (9), antibacterial (10) and antioxidant (11-13) properties of *S. reuterana* aerial parts. In an comparative study, Esmaeili *et al.* reported that methanol extract of *S. reuterana* had the highest DPPH free radical scavenging activity ( $IC_{50}$ ;  $15.1 \pm 1.00 \mu\text{g mL}^{-1}$ ) and ferric reducing power (FRAP value;  $0.34 \pm 0.01$ ), among the other tested *Salvia* species (11). Methanol extract of the *S. reuterana* flowers has also been reported to possess higher free radical scavenging activity with ( $IC_{50}$ ;  $77.6 \mu\text{g mL}^{-1}$ ) in comparison with its leave extract ( $IC_{50}$ ;  $119.4 \mu\text{g mL}^{-1}$ ), in DPPH assay (12).

As a result of phytochemical studies on this plant aerial part, nine labdane diterpenoids with cytotoxic activity against HeLa and MCF-7 cell lines were isolated from its *n*-hexane extract (14, 15). There are also some reports on essential oil composition of *S. reuterana* from different regions of Iran (10, 12, 16, 17). Regarding the results of previous studies on considerable antioxidant activity of this medicinal species, the aim of the present research was the isolation and identification of the compounds involved in free radical scavenging activity of *S. reuterana*.

## Experimental

### *Plant material*

The flowering aerial parts of *S. reuterana* were collected on Jun 2015 from the Khor region, Elburz province, Iran. A voucher specimen was deposited under the code 7045-TEH at the Herbarium of Faculty of Pharmacy, Tehran University of Medical Sciences, Tehran, Iran.

### *Extraction and fractionation*

The air dried and grinded plant aerial parts (400 g) were macerated with 80% methanol in water ( $5 \times 2.5 \text{ L}$ ). The total hydroalcoholic extract (82 g) was then dissolved in 500 mL of methanol-water mixture (8:2) and extracted by enough volumes of *n*-hexane and chloroform, successively, to get the *n*-hexane, chloroform and residual methanol-water (8:2) soluble (hydroalcoholic) main fractions.

### *Essential oil extraction*

Hydrodistillation method using Clevenger apparatus was used to essential extraction from 100 g of dried and comminuted plant material for 3 hours. The pale yellowish oil was dried over anhydrous sodium sulphate and kept at 4 °C until analysis.

### *DPPH free radical scavenging assay*

Antioxidant activity of the total extract, main fractions and essential oil of plant aerial parts were evaluated by DPPH (2, 2-diphenyl-1-picryl-hydrazyl) free radical scavenging assay method described by Sarker *et al.* (18). Briefly, twofold serial dilutions ( $1.0$  to  $3.9 \times 10^{-3} \text{ mg mL}^{-1}$ ) were made from samples, individually (each 2 mL). Two milliliter of freshly prepared DPPH (Sigma) solution ( $80 \mu\text{g mL}^{-1}$ ) was then added to each test tube. After 30 min, absorptions of the solutions were recorded at 517 nm using an Optizen 2120 UV PLUS spectrophotometer. Butylated hydroxy toluene (BHT) (Sigma), a commercial synthetic antioxidant, was also used as positive control. For each sample concentration causing a 50% reduction in absorption of DPPH solution ( $40 \mu\text{g mL}^{-1}$ ) was calculated as  $IC_{50}$ . The experiment was repeated three times and results were expressed as Mean  $\pm$  SD.

### *Isolation and purification of compounds*

Hydroalcoholic fraction with the highest free radical scavenging activity (Table 1) was subjected to further phytochemical analysis. A portion of this fraction (4 g) was chromatographed on a reversed-phase (RP18, mesh 230-400, Fluka) column and eluted with the gradient mixture of methanol in water (0.5-9.5 to 7:3) to get five fraction (M1-5). Compounds **1** (18 mg) and **2** (43 mg) were isolated from the fraction M3 (320 mg) on a Sephadex LH-20 column (Fluka) eluted by methanol:water (8:2) as solvent system. Column chromatography of the fraction M4 (147 mg) on a Sephadex LH-20 column with methanol resulted in the isolation of compound **3** (28 mg). Compound **4** (18 mg) was isolated from the fraction M5 (95 mg) through the reversed-phase column chromatography (methanol-water, 8:2). It was more purified on a Sephadex LH-20 column using methanol as eluent. All column chromatographies were



(1H, *br s*, H-2'), 6.94 (1H, *d*,  $J = 7.0$  Hz, H-5'), 6.82 (1H, *br s*, H-8), 6.74 (1H, *s*, H-3), 6.46 (1H, *br s*, H-6), 5.08 (1H, *d*,  $J = 7.5$  Hz, H-1''), 3.2-3.6 (6H, H-2''-6'').  $^{13}\text{C-NMR}$  (DMSO- $d_6$ , 125 MHz):  $\delta$  182.13 (C-4), 164.82 (C-2), 163.17 (C-7), 161.47 (C-5), 157.25 (C-9), 150.61 (C-4'), 146.15 (C-3'), 121.35 (C-1'), 119.51 (C-6'), 116.34 (C-5'), 113.60 (C-2'), 105.69 (C-10), 103.27 (C-3), 100.24 (C-1''), 100.14 (C-6), 95.00 (C-8), 77.34 (C-5''), 76.58 (C-3''), 73.37 (C-2''), 69.88 (C-4''), 60.93 (C-6'') (20, 21).

*Rosmarinic acid* ( $\alpha$ -O-caffeoyl-3,4-dihydroxyphenyllactic acid) (**3**);  $^1\text{H-NMR}$  (DMSO- $d_6$ , 500 MHz):  $\delta$  7.36 (1H, *d*,  $J = 16.1$  Hz, H-7), 7.06 (1H, *br s*, H-2), 6.89 (1H, *br d*,  $J = 6.5$ , H-6), 6.74 (1H, *d*,  $J = 6.5$  Hz, H-5), 6.68 (1H, *br s*, H-2'), 6.60 (1H, *d*,  $J = 7.0$  Hz, H-5'), 6.48 (1H, *br d*,  $J = 7.0$  Hz, H-6'), 6.17 (1H, *d*,  $J = 16.1$  Hz, H-8), 4.86 (1H, *d*,  $J = 8.5$ , H-8'), 3.02 (1H, *br d*,  $J = 13.5$  Hz, H-7'b), 2.75 (1H, *dd*,  $J = 13.5, 11$  Hz, H-7'a).  $^{13}\text{C-NMR}$  (DMSO- $d_6$ , 125 MHz):  $\delta$  172.97 (C-9'), 166.34 (C-9), 149.04 (C-4), 146.17 (C-7), 145.06 (C-3), 144.45 (C-3'), 143.69 (C-4'), 129.81 (C-1'), 125.25 (C-1), 120.81 (C-6), 119.59 (C-6'), 116.78 (C-2'), 116.05 (C-5), 115.49 (C-5'), 115.04 (C-2), 114.54 (C-8), 75.97 (C-8'), 37.17 (C-7') (22).

*Luteolin* (5,7,3',4'-Tetrahydroxyflavone) (**4**);  $^1\text{H-NMR}$  (DMSO- $d_6$ , 500 MHz):  $\delta$  7.42 (1H, *br d*,  $J = 8$  Hz, H-6'), 7.39 (1H, *br s*, H-2') 6.89 (1H, *d*,  $J = 8$  Hz, H-5'), 6.66 (1H, *s*, H-3), 6.44 (1H, *br s*, H-8), 6.18 (1H, *br s*, H-6).  $^{13}\text{C-NMR}$  (DMSO- $d_6$ , 125 MHz):  $\delta$  181.61 (C-4), 164.12 (C-2), 163.86 (C-7), 161.45 (C-5), 157.26 (C-9), 149.68 (C-4'), 145.71 (C-3'), 121.46 (C-1'), 118.93 (C-6'), 116.26 (C-5'), 113.32 (C-2'), 103.65 (C-10), 102.82 (C-3), 98.79 (C-6), 93.80 (C-8) (23).

Free radical scavenging activities of the isolated compounds (**1-4**) were also assessed by DPPH test. As shown in table 1, among the isolated compounds, luteolin (**4**), rosmarinic acid (**3**) and luteolin-7-O- $\beta$ -D glucopyranoside (**2**) were found to have a potent free radical scavenging activity with  $\text{IC}_{50}$  values of  $5.1 \pm 0.6$ ,  $9.6 \pm 1.2$ ,  $17.3 \pm 2.1$   $\mu\text{g mL}^{-1}$ , higher than positive control, BHT ( $\text{IC}_{50}$ :  $21.3 \pm 1.9$   $\mu\text{g mL}^{-1}$ ). Thus, these compounds can be assumed as the major free radical scavengers present in *S. reuterana* aerial parts.

**Table 1.** Free radical scavenging activities of the essential oil, extract, fractions and isolated compounds from *S. reuterana*.

Samples	$\text{IC}_{50}$ value ( $\mu\text{g mL}^{-1}$ ) <sup>a</sup>
Essential oil	246.4 $\pm$ 5.1
Total extract	187.6 $\pm$ 3.5
<i>n</i> -Hexane fraction	825.1 $\pm$ 12.6
Chloroform fraction	682.5 $\pm$ 4.3
Hydroalcoholic fraction	112.6 $\pm$ 3.2
Apigenin-7-O-glucoside ( <b>1</b> )	34.2 $\pm$ 1.3
Luteolin-7-O-glucoside ( <b>2</b> )	17.3 $\pm$ 2.1
Rosmarinic acid ( <b>3</b> )	9.6 $\pm$ 1.2
Luteolin ( <b>4</b> )	5.1 $\pm$ 0.6
BHT (Positive control)	21.3 $\pm$ 1.9

Note: <sup>a</sup> Concentration providing 50% inhibition.

Previously, Farimani and Miran reported the isolation six labdane diterpenoids, namely, sclareol, 6b-hydroxysclareol, 14a-epoxysclareol, 14a-hydroxy-15-chlorosclareol, 14a-hydroxy-15-acetoxysclareol and 6b-hydroxy-14a-epoxysclareol, together with two new diterpenoids, 6 $\beta$ ,14 $\alpha$ -dihydroxy-15-acetoxysclareol and 14 $\alpha$ ,15- dihydroxy sclareol from the *n*-hexane extract of *S. reuterana* aerial parts (14, 15). To our knowledge, this is the first report of the isolation and structure elucidation of these phenolic derivatives (**1-4**) from the aerial parts of this medicinal species. These compounds, however, have been isolated from various other *Salvia* species (24).

Some biological activities are found in literature for compounds **1-4** (25-43). Apigenin-7-O- $\beta$ -D-glucopyranoside (**1**) has been reported for its anxiolytic (25), insulin mimetic (26), antioxidant (27) and hepatoprotective (28) effects. Luteolin (**4**) and its 7-O-glucopyranoside derivative (**2**) have shown anti-inflammatory (29), chemopreventive (30, 31), antioxidant (32) and  $\alpha$ -glucosidase inhibitory (33) activities. Moreover, the results of recent studies reported luteolin as a flavonoid with neuroprotective and anxiolytic effects (34, 35). Accordingly, apigenin-7-O- $\beta$ -D-glucopyranoside and luteolin with known anxiolytic activity may be involved in anxiolytic properties of *S. reuterana*, which has been previously documented by Rabbani *et al.* (25). Rosmarinic acid (**3**), which has also been reported as a chemotaxonomic marker of the subfamily Nepetoideae (36), is a caffeic acid derivative

with a range of health benefit properties such as antioxidant (37), anti-inflammatory (38), antinociceptive (38), hepatoprotective (39) and neuroprotective (40) effects. In 2002, Takeda *et al.* showed that rosmarinic acid (2 mg/kg, i.p.) produces antidepressant-like effect in the forced swimming test in mice (41). Further studies indicated that this antidepressant-like effect is driven at least in part through the proliferation of newborn cells located in the dentate gyrus of the hippocampus (42). Rosmarinic acid has also been found as compound with  $\alpha$ -amylase inhibitory activity (43). Combination of  $\alpha$ -amylase and  $\alpha$ -glucosidase inhibitory effects and insulin mimetic activity, reported from the isolated compounds may be contributed

to antidiabetic properties of *S. reuterana*, previously published by Eidi *et al.* (9).

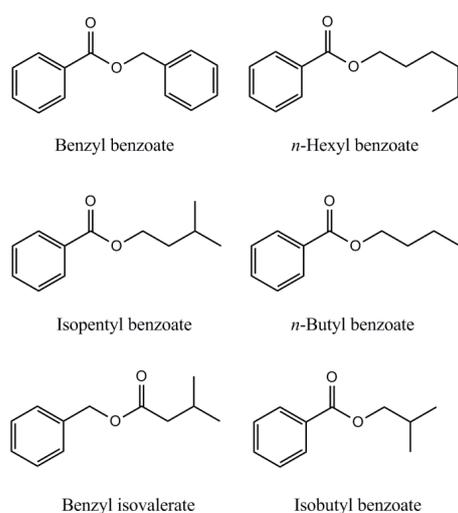
GC-MS analysis of the essential oil resulted in the identification of twenty four compounds, representing the 98.48% of the total oil. The essential oil was rich in non-terpene compounds (76.17%), mainly benzyl benzoate (26.64%), *n*-hexyl benzoate (22.99%) and *n*-hexyl isovalerate (6.04%) (Table 2). Essential oil extracted from *S. reuterana* aerial parts demonstrated notable DPPH free radical scavenging activity ( $IC_{50}$ :  $246.4 \pm 5.1 \mu\text{g mL}^{-1}$ ). However, the low yield of essential oil extraction (yield: 0.2% (v/w)) attenuates the importance of the plant essential oil in antioxidant properties of *S. reuterana*.

**Table 2.** Chemical composition of the essential oil of *S. reuterana* aerial parts.

No.	Compounds <sup>a</sup>	Rt <sup>b</sup>	RI <sup>c</sup>	%
1	<i>n</i> -hexyl acetate	8.27	1009	1.93
2	<i>n</i> -butyl isovalerate	8.75	1037	1.05
3	(E)- $\beta$ -ocimene	8.79	1046	0.68
4	isobutyric acid	10.07	1178	1.07
5	pentyl cyclopropane	10.79	1194	1.22
6	<i>n</i> -hexyl 2-methyl butyrate	11.09	1236	1.57
7	<i>n</i> -hexyl isovalerate	11.13	1245	6.04
8	isobutyl benzoate	12.11	1329	0.49
9	$\delta$ -elemene	12.24	1338	1.31
10	<i>n</i> -butyl benzoate	12.56	1356	4.45
11	benzyl isovalerate	12.75	1364	0.83
12	$\beta$ -elemene	12.79	1392	3.26
13	selin-4,7 (11)-diene	13.07	1412	1.49
14	isopentyl benzoate	13.17	1437	6.40
15	isolekene	13.62	1440	0.82
16	germacrene-D	13.67	1489	0.55
17	$\delta$ -selinene	13.73	1497	0.96
18	<i>n</i> -hexyl benzoate	14.43	1584	22.99
19	spathulenol	14.54	1588	1.07
20	$\beta$ -eudesmol	15.16	1654	3.14
21	benzyl benzoate	16.02	1767	26.64
22	sclareol oxide	17.02	1894	1.46
23	manoyl oxide	17.79	1932	0.69
24	sclareol	19.27	2218	8.37
	Monoterpene hydrocarbons			0.68
	Sesquiterpene hydrocarbons			6.90
	Oxygenated sesquiterpenes			4.21
	Diterpenes			10.52
	Non-terpenes			76.17
	Total identified			98.48

Note: <sup>a</sup> Identified compounds listed in order of elution from DB-5 column; <sup>b</sup> Retention times; <sup>c</sup> Retention indices to C<sub>8</sub>-C<sub>24</sub> *n*-alkanes on DB-5 column.

A review on the results of the present study and previous reports shows a variation in essential oil composition of *S. reuterana* aerial parts collected from different regions of Iran (10, 12, 16, 17). In an study by Fattahi *et al.* on essential oil analysis of seven wild population of *S. reuterana* from north and center of Iran,  $\alpha$ -gurjunene (5.4-13.7%),  $\beta$ -elemene (4.5-13.9%), germacrene D (2.6-7.2%), spathulenol (1.0-8.0%) and *n*-hexyl acetate (1.2-6.8%) were identified as major compounds (16). Benzyl benzoate, the main compound of our analyzed essential oil sample (26.64%), has been detected in the range of trace to 8.0% in former mentioned study (16). *n*-hexyl benzoate (22.99%), another main compound identified in the present study was not detected by Fattahi *et al.* in their examined essential oils of different *S. reuterana* populations (16). However, *n*-hexyl benzoate has been characterized at high amounts (17.0%) in essential oil of *S. reuterana* flowers, collected from Kashan region, center of Iran (12). Benzyl benzoate and *n*-hexyl benzoate have also been reported in essential oil of *Salvia multicaulis* Vahl aerial parts with relative percentages of 60.3 and 16.7 (44). Differences in climate conditions, as well as possible presence of chemotypes in various *S. reuterana* populations are the factors which could be assumed as responsible for the observed variations in essential oils composition (45). However, a comprehensive study using more advanced chromatographic and spectroscopic techniques is needed for the assessment of variations between essential oil contents of different populations of *S. reuterana*.



**Figure 2.** Structures of the aromatic compounds identified in essential oil of *S. reuterana* aerial parts.

## Conclusion

The results of present study verify that *S. reuterana* with its potent free radical scavenging flavonoid and caffeic acid derivatives content (1-4) can be considered as a valuable source of natural phenolic antioxidants. Literature review on biological activities of the isolated compounds also provides some molecular explanations for anxiolytic and antidiabetic properties, previously reported from *S. reuterana*. Moreover, interesting chemical composition of *S. reuterana* essential oils, which is dominated by non-terpenes compounds (76.17%), especially aromatic derivatives, make it an appropriate candidate for further studies.

## References

- (1) Aruoma OI. Free radicals, oxidative stress, and antioxidants in human health and disease. *J. Am. Oil Chem. Soc.* (1998) 75: 199-212.
- (2) Soobrattee MA, Neergheen VS, Luximon-Ramma A, Aruoma OI and Bahorun T. Phenolics as potential antioxidant therapeutic agents: mechanism and actions. *Mutat. Res.* (2005) 579: 200-13.
- (3) Masoud S, Alijanpoo B and Khayyami M. Contribution to cytology of genus *Salvia* L. (Lamiaceae) in Iran. *Caryologia.* (2010) 63: 405-10.
- (4) Jamzad Z. *Flora of Iran*. No.76: Lamiaceae. Publication of Research Institute of Forests and Rangelands, Tehran; (2012) 799-802.
- (5) Asghari G, Akbari M and Asadi-Samani M. Phytochemical analysis of some plants from Lamiaceae family frequently used in folk medicine in Aligudarz region of Lorestan province. *Marmara Pharm. J.* (2017) 21: 506-14.
- (6) Mohammadi H, Sajjadi SE, Noroozi M and Mirhosseini M. Collection and assessment of traditional medicinal plants used by the indigenous people of Dastena in Iran. *J. Herb. Med. Pharmacol.* (2016) 5: 54-60.
- (7) Rabbani M, Sajjadi S, Jafarian A and Vaseghi G. Anxiolytic effects of *Salvia reuterana* Boiss. on the elevated plus-maze model of anxiety in mice. *J. Ethnopharmacol.* (2005) 101: 100-3.
- (8) Vaseghi G, Andalib S, Rabbani M, Sajjadi S and Jafarian A. Hypnotic effect of *Salvia reuterana* Boiss for treatment of insomnia. *J. Med. Plants.* (2013) 12: 7-13.
- (9) Eidi A, Eidi M, Mozaffarian V and Rustaiyan A. Effect of *Salvia Reuterana* aerial parts on serum

- parameters in normal and streptozotocin-induced diabetic rats. *Health MED.* (2012) 6: 1199-206.
- (10) Esmacili A, Rustaiyan A, Nadimi M, Larujani K, Nadjafi F, Tabrizi L and Chalabian F. Chemical composition and antibacterial activity of essential oils from leaves, stems, and flowers of *Salvia reuterana* grown in Iran. *Chem. Nat. Compd.* (2008) 44:393-5.
- (11) Esmacili MA, Kanani MR and Sonboli A. *Salvia reuterana* extract prevents formation of advanced glycation end products: an in vitro study. *Iran. J. Pharm. Sci.* (2010) 6: 33-50.
- (12) Ghomi JS, Masoomi R, Kashi FJ and Batooli H. In vitro bioactivity of essential oils and methanol extracts of *Salvia reuterana* from Iran. *Nat. Prod. Com.* (2012) 7: 651-4.
- (13) Jeshvaghani ZA, Rahimmalek M, Talebi M and Goli SAH. Comparison of total phenolic content and antioxidant activity in different *Salvia* species using three model systems. *Ind. Crop. Prod.* (2015) 77: 409-14.
- (14) Moridi-Farimani M and Miran M. Labdane diterpenoids from *Salvia reuterana*. *Phytochemistry* (2014) 108: 264-9.
- (15) Moridi-Farimani M, Miran M and Nejad-Ebrahimi S. New Diterpenoids from the Aerial Parts of *Salvia reuterana*. *Iran. J. Pharm. Res.* (2018) 18: 406-11.
- (16) Fattahi B, Nazeri V, Kalantari S, Bonfill M and Fattahi M. Essential oil variation in wild-growing populations of *Salvia reuterana* Boiss. collected from Iran: Using GC-MS and multivariate analysis. *Ind. Crop. Prod.* (2016) 81: 180-90.
- (17) Mirza M and Sefidkon F. Essential oil composition of two *Salvia* species from Iran, *Salvia nemorosa* L. and *Salvia reuterana* Boiss. *Flav. Fragr. J.* (1999) 14: 230-2.
- (18) Sarker SD, Latif Z and Gray AI. *Natural products isolation*. 2nd Ed. Human Press Inc, New Jersey (2006) 20-1.
- (19) Adams RP. *Identification of Essential Oil Components by Gas Chromatography/Mass Spectrometry*. Allured Publishing Corporation, Carol Stream (2007).
- (20) Simaratanamongkol A, Umehara K, Noguchi H and Panichayupakaranant P. Identification of a new angiotensin-converting enzyme (ACE) inhibitor from Thai edible plants. *Food Chem.* (2014) 165: 92-7.
- (21) Orhan F, Bariş Ö, Yanmiş D, Bal T, Güvenalp Z and Güllüce M. Isolation of some luteolin derivatives from *Mentha longifolia* (L.) Hudson subsp. *longifolia* and determination of their genotoxic potencies. *Food Chem.* (2012) 135: 764-9.
- (22) Ha TJ, Lee JH, Lee MH, Lee BW, Kwon HS, Park CH, Shim K, Kim H, Back I and Jang DS. Isolation and identification of phenolic compounds from the seeds of *Perilla frutescens* (L.) and their inhibitory activities against  $\alpha$ -glucosidase and aldose reductase. *Food. Chem.* (2012) 135: 1397-403.
- (23) Loizzo MR, Said A, Tundis R, Rashed K, Statti GA, Hufner A and Menichini F. Inhibition of angiotensin converting enzyme (ACE) by flavonoids isolated from *Ailanthus excelsa* (Roxb)(Simaroubaceae). *Phytotherapy Res.* (2007) 21: 32-6.
- (24) Lu Y and Foo LY. Polyphenolics of *Salvia*-a review. *Phytochemistry* (2002) 59:117-40.
- (25) Kumar D and Bhat ZA. Apigenin 7-glucoside from *Stachys tibetica* Vatke and its anxiolytic effect in rats. *Phytomed.* (2014) 21: 1010-4.
- (26) Rao YK, Lee MJ, Chen K, Lee YC, Wu WS and Tzeng YM. Insulin-mimetic action of rhoifolin and cosmosiin isolated from *Citrus grandis* (L.) Osbeck leaves: enhanced adiponectin secretion and insulin receptor phosphorylation in 3T3-L1 cells. *Evid-Based. Compl. Alt. Med.* (2011) 1-9.
- (27) Benavente-Garcia O, Castillo J, Lorente J, Ortuno A and Del Rio J. Antioxidant activity of phenolics extracted from *Olea europaea* L. leaves. *Food Chem.* (2000) 68: 457-62.
- (28) Zheng Q, Sun X, Xu B, Li G and Song M. Mechanisms of apigenin-7-glucoside as a hepatoprotective agent. *Biomed. Environ. Sci.* (2005) 18: 65-70.
- (29) Park CM and Song Y-S. Luteolin and luteolin-7-O-glucoside inhibit lipopolysaccharide-induced inflammatory responses through modulation of NF- $\kappa$ B/AP-1/PI3K-Akt signaling cascades in RAW 264.7 cells. *Nutr. Res. Pract.* (2013) 7 :423-9.
- (30) Manju V and Nalini N. Chemopreventive potential of luteolin during colon carcinogenesis induced by 1, 2-dimethylhydrazine. *Ital. J. Biochem.* (2005) 54: 268-75.
- (31) Baskar AA, Ignacimuthu S, Michael GP and Al-Numair KS. Cancer chemopreventive potential of luteolin-7-O-glucoside isolated from *Ophiorrhiza mungos* Linn. *Nutr. Cancer.* (2011) 63: 130-8.
- (32) Song YS and Park CM. Luteolin and luteolin-7-O-glucoside strengthen antioxidative potential through the modulation of Nrf2/MAPK mediated HO-1 signaling cascade in RAW 264.7 cells. *Food Chem. Toxicol.* (2014) 65: 70-5.
- (33) Kim J-S, Kwon C-S and SoN KH. Inhibition of alpha-glucosidase and amylase by luteolin, a flavonoid. *Biosci. Biotechnol. Biochem.* (2000) 64: 2458-61.
- (34) Nabavi SF, Braidly N, Gortzi O, Sobarzo-Sanchez E, Daglia M, Skalicka-Woźniak K and Nabavi SM. Luteolin as an anti-inflammatory and neuroprotective agent: A brief review. *Brain Res. Bull.* (2015) 119: 1-11.

- (35) Coleta M, Campos MG, Cotrim MD, de-Lima TCM and da-Cunha AP. Assessment of luteolin (3', 4', 5, 7-tetrahydroxyflavone) neuropharmacological activity. *Behav. Brain Res.* (2008) 189: 75-82.
- (36) Janicsák G, Máthé I, Miklóssy-Vári V and Blunden G. Comparative studies of the rosmarinic and caffeic acid contents of *Lamiaceae* species. *Biochem. Syst. Ecol.* (1999) 27: 733-8.
- (37) Chen JH and Ho CT. Antioxidant activities of caffeic acid and its related hydroxycinnamic acid compounds. *J. Agric. Food Chem.* (1997) 45: 2374-8.
- (38) Boonyarikpunchai W, Sukrong S and Towiwat P. Antinociceptive and anti-inflammatory effects of rosmarinic acid isolated from *Thunbergia laurifolia* Lindl. *Pharmacol. Biochem. Behav.* (2014) 124: 67-73.
- (39) Domitrović R, Škoda M, Marchesi VV, Cvijanović O, Pugel EP and Štefan MB. Rosmarinic acid ameliorates acute liver damage and fibrogenesis in carbon tetrachloride-intoxicated mice. *Food Chem. Toxicol.* (2013) 51: 370-8.
- (40) Braidy N, Matin A, Rossi F, Chinain M, Laurent D and Guillemin G. Neuroprotective effects of rosmarinic acid on ciguatoxin in primary human neurons. *Neurotox. Res.* (2014) 25: 226-34.
- (41) Takeda H, Tsuji M, Inazu M, Egashira T and Matsumiya T. Rosmarinic acid and caffeic acid produce antidepressive-like effect in the forced swimming test in mice. *Europ. J. Pharmacol.* (2002) 449: 261-7.
- (42) Ito N, Yabe T, Gamo Y, Nagai T, Oikawa T, Yamada H and Hanawa T. Rosmarinic acid from *Perillae Herba* produces an antidepressant-like effect in mice through cell proliferation in the hippocampus. *Biol. Pharma. Bull.* (2008) 31: 1376-80.
- (43) McCue PP and Shetty K. Inhibitory effects of rosmarinic acid extracts on porcine pancreatic amylase *in-vitro*. *Asia. Pac. Clin. Nutr.* (2004) 13: 101-106.
- (44) Mojtaba T, Reza GH, Borzo S, Shiva N and Esmail S. *In-vitro* antibacterial and antifungal activity of *Salvia multicaulis*. *J. Essent. oil bearing plants.* (2011) 14: 255-9.
- (45) Baser KHC and Buchbauer G. *Handbook of Essential Oils: Science, Technology, and Applications*. CRC Press, Florida (2009) 60-70.
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